



Research Article

# Methyltransferase-like 14 promotes the tumorigenesis and proliferation of pancreatic cancer cells through myc proto-oncogene signaling pathway

Junru Li MM<sup>#1</sup>, Peng Wang MM<sup>#1</sup>, Fei Liu MM<sup>1</sup>, Yuanyuan Li MB<sup>1</sup>, Youyou Wu MM<sup>1</sup>, Fengbo Wang MM<sup>1</sup>, Jundong Du MD<sup>1</sup>

<sup>1</sup>Department of General Surgery, Jincheng General Hospital, Jincheng, China.

<sup>#</sup>Contributed equally to this work



**\*Corresponding author:**

Jundong Du,  
Department of General Surgery,  
Jincheng General Hospital,  
Jincheng, China.

18518073878@163.com

Received: 17 June 2024

Accepted: 23 September 2024

Published: 26 November 2024

**DOI**

10.25259/Cytojournal\_105\_2024

**Quick Response Code:**



## ABSTRACT

**Objective:** Pancreatic cancer is characterized by low survival rate and rapid deterioration. Methyltransferase-like 14 (METTL14), as N6-methyladenosine (m6A) methyltransferase, is closely related to tumor progression. The purpose of this study is to look into how METTL14 affects pancreatic cancer tumorigenesis, cell division, and apoptosis.

**Material and Methods:** We examined and contrasted the levels of METTL14 protein and messenger RNA expression in human pancreatic ductal cells and human pancreatic cancer cells. After silencing or upregulating METTL14, the proliferative ability, migration ability, and cell apoptosis of pancreatic tumor cells was detected by colony-forming assay, wound scratch healing assay, cell counting kit 8 assay, and terminal deoxynucleotidyl transferase-mediated 2'-deoxyuridine 5'-triphosphate-biotin nick end labeling assay. Following the use of c-Myc inhibitor (10058-F4), western blot analysis was carried out to investigate the key factor expression and c-Myc signaling pathway activation status.

**Results:** METTL14 was preferentially expressed in human pancreatic cancer cells PANC-1 and SW1990 than in human normal pancreatic duct cells human pancreatic nestin-expressing cells (HPNE) ( $P < 0.001$ ). Overexpression of METTL14 increased the tumorigenic and proliferative ability of pancreatic cancer cells. Overexpression of METTL14 decreased apoptosis rate. Western blot assay showed that nucleus  $\beta$ -catenin increased when METTL14 was overexpressed, and nucleus  $\beta$ -catenin decreased when METTL14 was silenced in PANC-1 cell ( $P < 0.01$ ). The protein expression of other key factors, such as c-Myc, matrix metalloproteinase (MMP)-9, and MMP-2, were also affected. The use of c-Myc inhibitor (10058-F4) on the basis of OE-METTL14 reversed the effect of the overexpression of METTL14 on promoting the tumorigenesis and cell proliferation of pancreatic cancer cell lines PANC-1 and SW1990.

**Conclusion:** METTL14 promoted the tumorigenesis and proliferation of pancreatic cancer cells by the c-Myc signaling pathway.

**Keywords:** Methyltransferase-like 14, Myc proto-oncogene, Pancreatic cancer, Tumorigenesis, Proliferation

## INTRODUCTION

Pancreatic cancer remains one of the most lethal malignancies and is characterized by an extremely low 5-year survival rate due to its aggressive nature and late diagnosis.<sup>[1,2]</sup> Common clinical treatment for pancreatic cancer is a combination of surgical resection and adjuvant chemotherapy.<sup>[3-5]</sup> Surgical treatment of pancreatic cancer has made significant advancements.

Recently, three recent surgical approaches have improved the prognosis of patients with non-metastatic operable pancreatic cancer.<sup>[6]</sup> Advancements in modern imaging, radiotherapy techniques, and application of chemotherapy have significantly improved the control rates of pancreatic cancer. Despite these advances, the incidence of pancreatic cancer still increases. Treatment of pancreatic cancer can be further improved.<sup>[7]</sup>

Methyltransferase-like 14 (METTL14) usually acts as a N<sup>6</sup>-methyladenosine (m<sup>6</sup>A) methyltransferase in tumors and is closely related to tumor progression through methylation.<sup>[8-10]</sup> METTL14 may also affect the development of cancer by participating in the stability of RNAs.<sup>[11,12]</sup> Increasing studies have found that METTL14 could affect tumor progression through multiple mechanisms.<sup>[13]</sup> Wang *et al.*<sup>[14]</sup> illustrated that METTL14 could reduce PERP expression and promote tumor proliferation and cancer metastasis in pancreatic cancer. Chen *et al.* found that METTL14 deletion can remarkably reduce the lymphatic metastasis of pancreatic cancer.<sup>[15]</sup> Numerous canonical roles for the wingless-related integration site (Wnt)/ $\beta$ -catenin signaling system include tumor metabolism, embryonic development, cell fate determination, and the epithelial-mesenchymal transition. The Wnt pathway's activation encourages  $\beta$ -catenin to build up in the cytoplasm and then go into the cell nucleus. In nucleus,  $\beta$ -catenin could act on T cell factor/lymphoid enhancer factor (TCF/LEF) promoters and regulate downstream genes of the Wnt pathway at the transcriptional level.<sup>[16]</sup> Moreover, c-Myc is commonly upregulated in pancreatic cancer and associated with its aggressive behavior.<sup>[17]</sup> As such, c-Myc has proved to be a target of METTL14.<sup>[18-20]</sup> This potential crosstalk among METTL14, c-Myc, and the Wnt/ $\beta$ -catenin pathway suggests a complex regulatory network that enhances pancreatic cancer cell proliferation and tumor progression.

Human pancreatic ductal cells and human pancreatic cancer cells were cultured *in vitro* to compare METTL14 level between pancreatic cancer and normal cells to verify our hypothesis and explore the function of METTL14 in pancreatic cancer. METTL14 expression in cells was regulated, and the cell proliferation rate, c-Myc expression level, and key protein activation status of the Wnt signaling pathway were compared. The c-Myc inhibitor 10058-F4 was used to determine whether METTL14 targets c-Myc in pancreatic cancer.

## MATERIAL AND METHODS

### Pancreatic cell culture

Normal pancreatic ductal cell line (HPNE; Cat No. iCell-h102, iCell Bioscience Inc., Shanghai, China) and pancreatic cancer cell lines (PANC-1, Cat No. iCell-h172, iCell Bioscience Inc.;

SW1990, Cat No. iCell-h203, iCell Bioscience Inc.) were purchased from iCELL Biotechnology Company (Shanghai, China). All the three cell lines were cultured in 1640 medium (Cat No. iCell-0002, iCell Bioscience Inc, Shanghai, China) with 5% carbon dioxide (CO<sub>2</sub>) at 37°C. The culture medium contained 10% fetal bovine serum (C0226, Beyotime, Shanghai, China) and 5% streptomycin/penicillin (15140122, Thermo Fisher Scientific, Waltham, MA, USA). Short tandem repeat testing was applied to authenticate the three cell lines, and mycoplasma testing was performed before using the cells to ensure that they are not contaminated.

### Cell transfection and cell treatment

Plasmids encoding small interference RNA (si-RNA) against METTL14 (si-METTL14) or negative control (si-NC) were transfected into si-METTL14 group (si-METTL14 #1: 5'-GGAUGAGUAAUAGCUAAAUC-3', 5'-UUUAGCUAUAACUCAUCCUU-3'; si-METTL14 #2: 5'-GGAUGAUUAUGAAGUUAGA-3', 5'-UAACUUCAUAAUAUCAUCCCA-3'; si-METTL14 #3: 5'-GACUGGUUGUACAGAAGAAAU-3', 5'-UUCUUCUGUACAACCAGUCA-3') or si-NC group (5'-UUCUCCGAACGUGUCACGUTT-3' and 5'-ACGUGACACGUUCGGAGAATT-3') (GenePharma, Shanghai, China). The three si-RNAs were tested in our preliminary experiments, and the most effective one (si-METTL14 #1) was transfected in the si-METTL14 group. Plasmids encoding METTL14 (OE-METTL14, 5'-GTGCAGAAGGGACTAGGCAG-3', 5'-GGGCACCCACGTAATAGACC-3') or blank plasmids (OE-NC) were transfected in the OE-METTL14 group or OE-NC group. All plasmids were designed by GeneChem (Shanghai, China), and Lipofectamine™ 3000 was used for transfection (L3000001, Invitrogen, CA, USA). The OE-METTL14+10058-F4 group was treated with c-Myc inhibitor 10058-F4 (Cat No. HY-12702, MedChemExpress, Monmouth Country, NJ, USA) after transfection of OE-METTL14.

### Quantitative real-time reverse transcription polymerase chain reaction (qRT-PCR) assay

TRNzol Universal RNA simple Total RNA Kit (Cat No. DP424, Tiangen, Beijing, China) was used in total RNA extraction. Reverse transcription was conducted using FastQuant complementary DNA Synthesis Kit (Cat No. KR116, Tiangen). All measurements were analyzed using 2<sup>- $\Delta\Delta$ CT</sup> method. The sequence is shown in Table 1.

### Western blot assay

The collected cells were mixed well with Radio Immunoprecipitation Assay (Cat No. P0013, Beyotime, Shanghai, China). The protein concentrations were measured

**Table 1:** The primer sequences.

Primer	Primer sequences (5'-3')
human METTL14 forward	AGTTGGAACATGGATAGC
human METTL14 reverse	CTTCAACAATTATCGAGG
mouse $\beta$ -catenin forward	TGACACCTCCCAAGTCCTTT
mouse $\beta$ -catenin reverse	TTGCATACTGCCCGTCAAT
mouse MMP-9 forward	CCTGGAACACACGACATCTTC
mouse MMP-9 reverse	TGGAAACTCACACGCCAGAA
mouse MMP-2 forward	ACCCAGATGTGGCCAACTAC
mouse MMP-2 reverse	TACTTTTAAGGCCGAGCAA
mouse c-Myc forward	GCGACTCTGAAGAAGAGCAAG
mouse c-Myc reverse	GCCTCGGGATGGAGATGAG
GAPDH forward	CAATGACCCCTTCATTGACC
GAPDH reverse	GATCTCGCTCCTGGAAGATG

METTL14: Methyltransferase-like 14, c-Myc: Myc proto-oncogene, MMP: Matrix metalloproteinase, GAPDH: Glyceraldehyde-3-phosphate dehydrogenase, A; Adenosine, C; Cytosine, G; Guanine, T; Thymine.

using the bicinchoninic acid assay (BCA) kit (A045-4, Nanjing Institute of Bioengineering, Jiangsu, China) after the supernatant was collected. The soluble protein was denatured by heating it at 100°C for 5 min after being added at a ratio of one to ten to the sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) loading buffer (BL529B, Biosharp Life Science, Hefei, Anhui, China). 15% SDS-PAGE (BL522A, Biosharp Life Science, Hefei, Anhui, China) electrophoresis was used to separate the denatured proteins. Targeted primary antibodies were incubated at 4°C while the separated proteins were transferred to the membranes (IPVH00010, Millipore Sigma, Billerica, MA, USA). All primary antibodies used in the research were listed as follows: Polyclonal METTL14 (1:1000 dilution; ab309096) and c-Myc (1:1000 dilution; ab185656) were obtained from Abcam (Cambridge, MA, USA).  $\beta$ -Catenin (1:2000 dilution, #8480), matrix metalloproteinase (MMP)-9 (1:2000 dilution, #3852), MMP-2 (1:2000 dilution, #40994), and Histone H3 (1:3000 dilution, #4499) were purchased from CST (Danvers, MA, USA) anti-glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (1:1000 dilution; PA1-987) was purchased from Thermo Fisher Scientific (Waltham, MA, USA). Secondary antibody (1:2000 dilution; horseradish-peroxidase goat anti-rabbit immunoglobulin G; ab7090, Abcam, Cambridge, MA, USA) was incubated at room temperature. Protein bands were displayed with enhanced chemiluminescence (ECL) kits (BL520B, Biosharp Life Science, Hefei, Anhui, China) and Image Quant LAS4000 (GE Healthcare, Chicago, IL, USA) and analyzed by ImageJ software (version 2.0, LOCI, University of Wisconsin, Madison, Wisconsin, USA). Mean gray value of each protein band was recorded as raw

measurement. The background signal for each band was subtracted to correct for non-specific staining. The intensity of each band was then normalized to a loading control (GAPDH or histone H3) to account for variations in protein loading or transfer efficiency.

### Cell counting kit 8 (CCK8) assay

Cells were seeded in equal numbers into 96-well plates. Each detection hole contained  $1.5 \times 10^5$  cells. Cell proliferation was assessed using the CCK8 (Cat No. G4103-1ML, Servicebio, Wuhan, Hubei, China) at 24-, 48-, 72-, and 96-h post-seeding. 15  $\mu$ L of CCK-8 solution was applied to each well at the designated time and left for 2 h. Absorbance at 450 nm was recorded by a DR-200Bs reader (Diatek, Jiangsu, China).

### Colony-forming assay

Cells were digested with pancreatic protein (Cat No. T1300, Solarbio, Beijing, China). A total of  $1.5 \times 10^3$  cells were added to each well and cultured in an incubator at 37°C and 5% CO<sub>2</sub>. Each well was stained with 0.1 % crystal violet (Cat No. C0121, Beyotime) dye. The number of clones was analyzed and counted using ImageJ software.

### Wound scratch healing assay

In brief,  $1.5 \times 10^5$  cells were seeded into a 12-well culture plate. Once the cells reached 100% confluency, a wound was created using a 20  $\mu$ L pipette tip. Images were captured using an optical microscope at  $\times 200$  magnification (#ECLIPSE Ts2, Nikon, Tokyo, Japan) at the beginning of the assay and after 24 h of treatment. Migration ratio was then calculated using ImageJ software.

### TdT-mediated dUTP-biotin nick end labeling (TUNEL) assay

The cells were fixed and permeabilized by 30 min of treatment with 4% paraformaldehyde and subsequent 5 min of treatment with 0.3% Triton X-100 (P0096, Beyotime, Shanghai, China), respectively. In the darkness at 37°C, the cells were incubated with terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) solution (C1089, Beyotime, Shanghai, China) for 60 min and 4',6-Diamidino-2'-phenylindole (DAPI, BS097, Biosharp Life Science, Hefei, Anhui, China) for 15 min. DAPI is used to stain the nucleus. Four random fields were randomly selected for counting apoptotic cells using a fluorescence microscope (CKX53, OLYMPUS, Japan).

### In vivo tumor growth assay

Ten six-week-old male Balb/c nude mice ( $18 \pm 2$  g) were obtained from the Jincheng General Hospital Animal

Center (Jincheng, Shanxi, China). All animal experiments were conducted following the ethical guidelines approved by the Ethics Committee of Jincheng General Hospital (Approval No.:2024072301). PANC-1 cells ( $1 \times 10^7$ ) stably transfected with either the METTL14 knockdown vector or a control no-load lentivirus vector were injected into the right flanks of the mice. The mice were divided into shCtrl and shMETTL14 groups. After 5 weeks, the mice were euthanized by neck dislocation. The xenografted tumors were harvested for hematoxylin and eosin (HE) staining and immunohistochemical (IHC) analysis. Tumor volumes were calculated using the formula: volume = (length  $\times$  width<sup>2</sup>)/2.

### Lentivirus transfection

Lentiviral vectors containing shCtrl (pLKO.1) and shMETTL14 (5'-ACCTCGCCGTGTTAAATAGCAAAGATCAAGAGATCTTTGCTATTTAACACGGCTT-3') were constructed by GenePharma (Shanghai, China) and utilized for cell transfection. PANC-1 cells were transfected for 48 h. The cells were cultured in medium containing 5  $\mu$ g/mL puromycin (P8833, Sigma-Aldrich, Billerica, MA, USA) for 2 weeks to select for stably transfected cells. The verified pools of knockdown cells were then utilized for subsequent experiments.

### HE staining and IHC analysis

The tumor tissues were sectioned to a thickness of 4  $\mu$ m after being fixed in paraffin. The tissue samples were then stained with HE or incubated with the specified primary antibodies using the Elivision™ plus polymer horseradish peroxidase immunohistochemistry kit (D41-18, Maxim, Fujian, China).

For HE staining, hematoxylin-eosin stain kit (G1120, Solarbio, Beijing, China) was used.

For IHC assay, the tumor sections incubated with the primary antibody Ki-67 (1:1000 dilution, #9449S, CST, Danvers, MA, USA).

The staining images were observed by microscope (CKX53, OLYMPUS, Tokyo, Japan), and lung metastasis was quantified. Lung metastasis were evaluated by HE staining. Ki-67 was statistically analyzed by ImageJ software (v1.8.0.345, National Institutes of Health, Bethesda, MD, USA).

### Immunofluorescence staining

Immunofluorescence was used to locate  $\beta$ -catenin. After being fixed in 4% paraformaldehyde, the cells were injected into phosphate-buffered saline for 10 min with 0.1% Triton X-100. The cells were sealed with 2% bovine serum albumin for 15 min. Primary antibody  $\beta$ -catenin (1:200 dilution, #8480T, CST, Danvers, MA, USA) was incubated at 4°C

overnight. The nuclear were stained with DAPI (C1002, Beyotime Biotechnology, Shanghai, China). Finally, laser scanning confocal microscopy (LSM900, Zeiss, Oberkochen, Baden-Württemberg, Germany) was used for imaging.

### Statistical analysis

The Statistical Package for the Social Sciences 20.0 software was used for data analysis (Analysis Software Co., Ltd., Shanghai, China). All the data were presented as mean  $\pm$  standard deviation and assessed by independent two-tailed Student's *t*-test. Multiple comparisons between multiple sets of data were conducted using one-way analysis of variance with Tukey's test for *post hoc* analysis. Data analysis was performed using GraphPad 8.0.2 software (GraphPad Software, La Jolla, CA, USA). Statistical significance was considered at  $P < 0.05$ .

## RESULTS

### Expression of METTL14 in different cells

The protein levels of METTL14 in PANC-1 ( $P < 0.001$ ) and SW1990 ( $P < 0.001$ ) were higher than that in HPNE [Figure 1a and b]. Furthermore, METTL14 messenger (mRNA) had higher expression in PANC-1 [Figure 1c], ( $P < 0.001$ ) and SW1990 [Figure 1c], ( $P < 0.001$ ) than in HPNE.

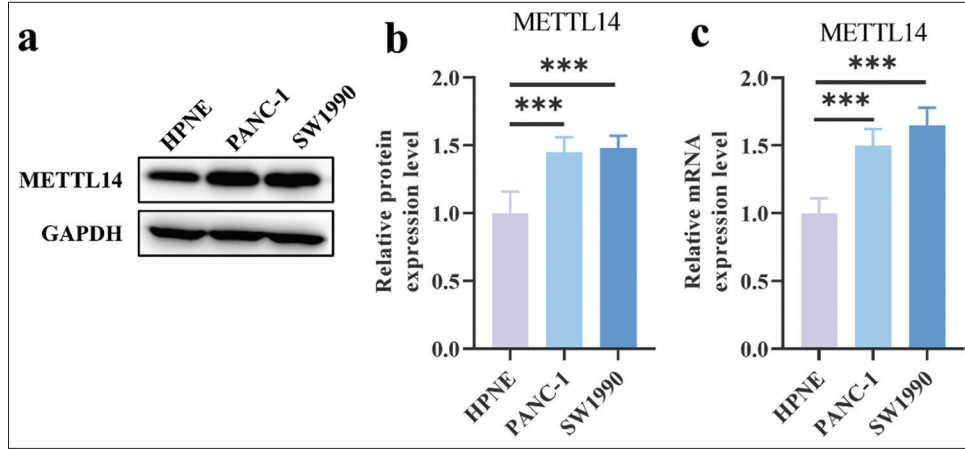
### METTL14 promoted the proliferation and migration of pancreatic cancer cells

The CCK8 assay results in transfected PANC-1 cells [Figure 2a] and SW1990 cells [Figure 2b] demonstrated that the cell viability increased in OE-METTL14-transfected cells ( $P < 0.01$ ) but decreased in si-METTL14-transfected cells ( $P < 0.01$ ). Figure 2c-e showed that the colony formation number of OE-METTL14-transfected cells ( $P < 0.001$ ) increased; this number in si-METTL14-transfected cells ( $P < 0.001$ ) decreased. In the results of wound scratch healing assay [Figure 2f-h], the cancer cell migration ability increased in OE-METTL14-transfected cells ( $P < 0.001$ ) but decreased in si-METTL14-transfected cells ( $P < 0.001$ ). In Figure 2i-k, OE-METTL14-transfected groups ( $P < 0.01$ ) had less apoptotic pancreatic cancer cells than OE-NC groups. The si-NC groups had less apoptotic pancreatic cancer cells than the si-METTL14-transfected group ( $P < 0.01$ ). These results showed that METTL14 promoted the tumor proliferation and migration of pancreatic cancer.

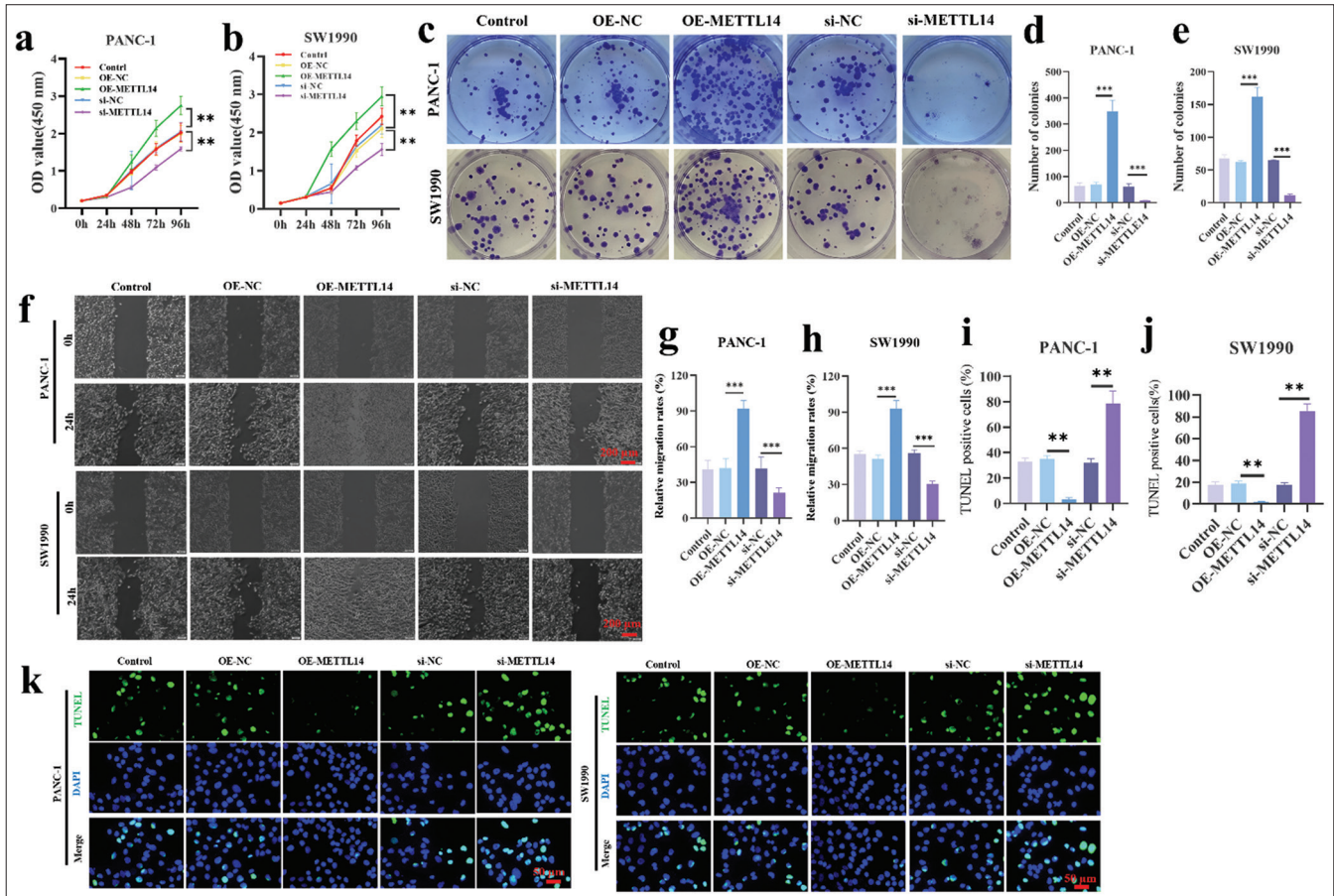
### METTL14 promoted c-Myc expression and nucleus $\beta$ -catenin expression in pancreatic cancer cell lines

The protein expression of key factors such as c-Myc,  $\beta$ -catenin, MMP-9, and MMP-2 was explored. Figure 3a-g showed that the expression of c-Myc, MMP-2, and MMP-

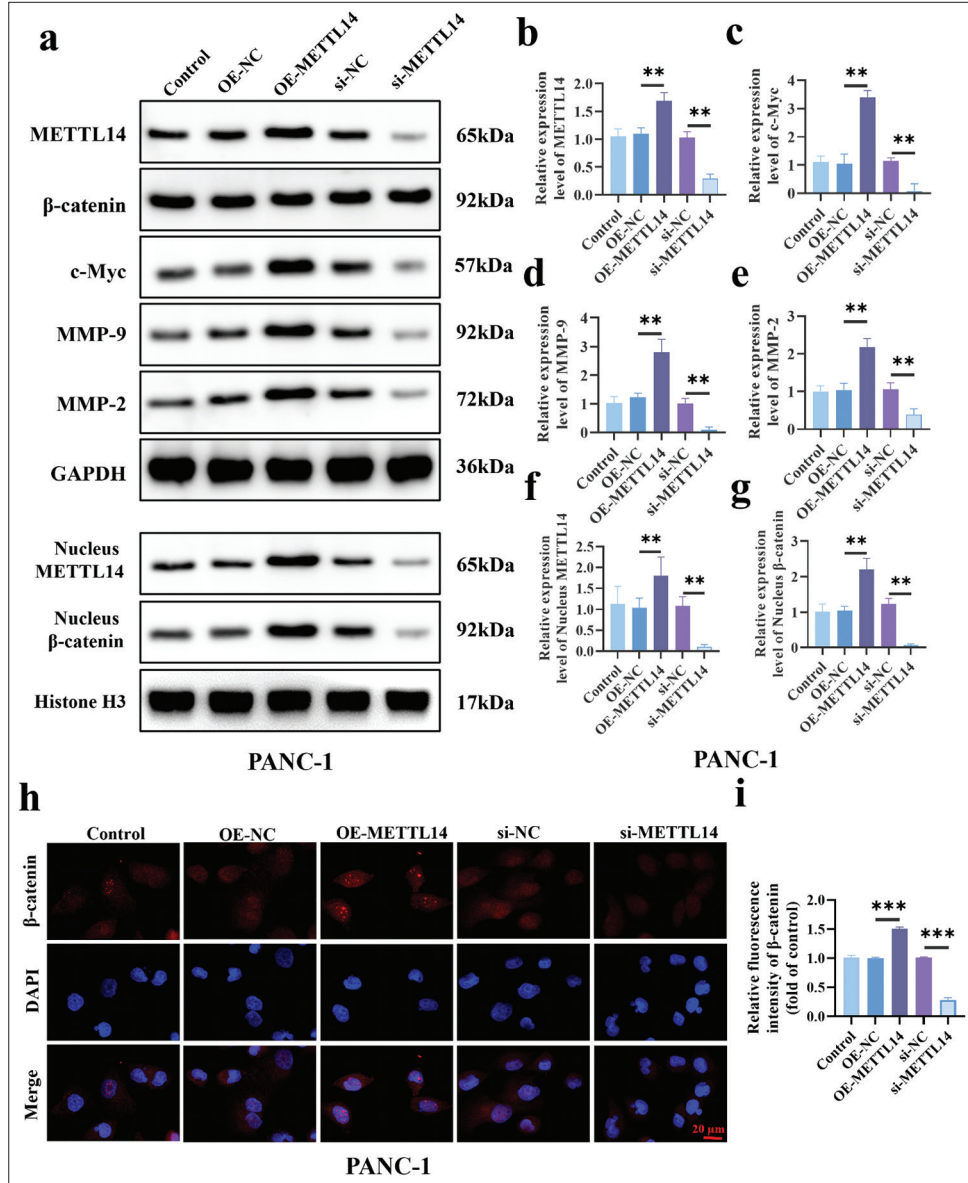




**Figure 1:** Expression of METTL14 in different cells. (a) Representative images of Western blot results and protein level of METTL14. (b) Quantitative analysis of relative protein expression of METTL14. (c) messenger RNA expression of METTL14.  $n = 3$ ,  $***P < 0.001$ . (METTL14: Methyltransferase-like 14, GAPDH: Glyceraldehyde-3-phosphate dehydrogenase).



**Figure 2:** METTL14 promoted the proliferation and migration of pancreatic cancer cells. (a and b) CCK8 assay after silencing and overexpressing METTL14. (c-e) colony formation assay after silencing and overexpressing METTL14 (crystal violet staining). (f-h) wound scratch healing assay after silencing and overexpressing METTL14. Objective: 400×. (i-k) TUNEL assay after silencing and overexpressing METTL14s. Objective: 400×.  $n = 3$ ,  $**P < 0.01$ ,  $***P < 0.001$ . (TUNEL: Terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling, DAPI: 4',6-diamidino-2-phenylindole, METTL14: Methyltransferase-like 14).



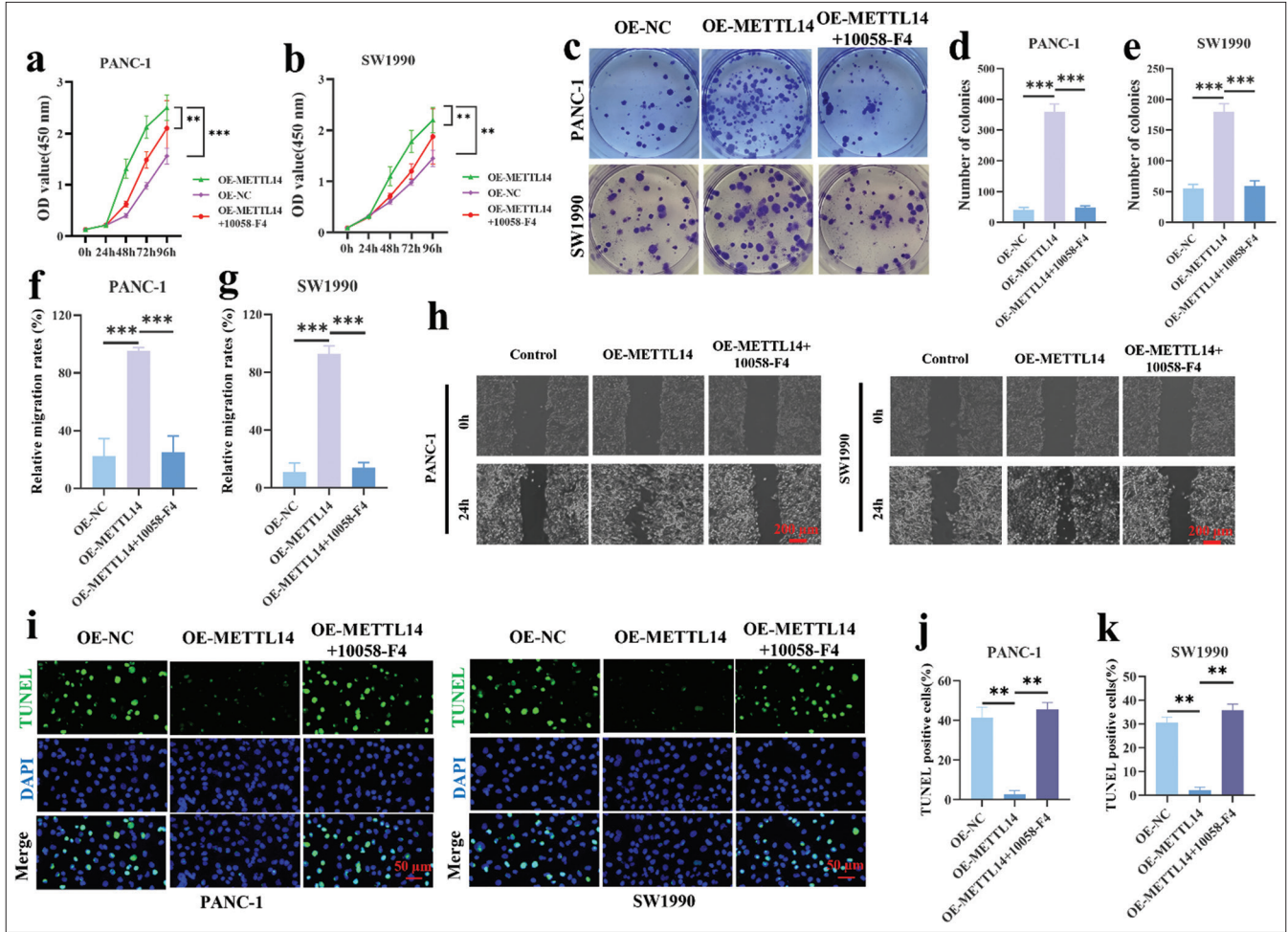
**Figure 3:** METTL14 promotes c-Myc expression and nucleus β-catenin expression *in vitro*. (a) Western blot results of METTL14, c-Myc, β-catenin, MMP-9, and MMP-2 in PANC-1. (b-g) Relative protein expression of METTL14, c-Myc, β-catenin, MMP-9, and MMP-2 in PANC-1. (h-i) β-catenin immunofluorescence assay. Objective: 200×. n = 3, \*\*P < 0.01, \*\*\*P < 0.001. (c-Myc: Myc proto-oncogene, MMP: Matrix metalloproteinase, METTL14, methyltransferase-like 14, DAPI: 4',6'-diamidino-2-phenylindole, GAPDH: Glyceraldehyde-3-phosphate dehydrogenase).

9 increased iafter METTL14 treatment and decreased after silencing METTL14. Interestingly, total expression of β-catenin remained unchanged in the OE-METTL14 group and si-METTL14 group, while nucleus β-catenin protein expression was increased after METTL14 overexpression and decreased after silencing METTL14 ( $P < 0.01$ ). The results of β-catenin immunofluorescence staining are shown in Figure 3h and i. The positive β-catenin signal was strong in the OE-METTL14 group but weak in the si-METTL14

group. All these results indicated that METTL4 could promote the protein expression of c-Myc, MMP-9, and MMP-2, but not β-catenin. METTL4 may induce the movement of β-catenin into the nucleus and activate β-catenin signaling.

#### Effect of c-Myc inhibitor (10058-F4) on METTL14 overexpressed pancreatic cancer cells

The CCK8 assay [Figure 4a] showed that PANC-1 cells in the OE-METTL14 group proliferated more than cells



**Figure 4:** Effect of c-Myc inhibitor (10058-F4) on METTL14 overexpressed pancreatic cancer cells. (a and b) The 10058-F4 treatment of METTL14 overexpressed cells was analyzed by CCK-8 assay. (c-e) The 10058-F4 treatment of METTL14 overexpressed cells was analyzed by colony formation assay (crystal violet staining). (f-h) The 10058-F4 treatment of METTL14 overexpressed cells was analyzed by wound scratch healing assay. Objective: 400 $\times$ . (i-k) The 10058-F4 treatment of METTL14 overexpressed cells was analyzed by TUNEL assay. Objective: 400 $\times$ .  $n = 3$ ,  $**P < 0.01$ ,  $***P < 0.001$ . (METTL14: Methyltransferase-like 14, TUNEL: Terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling, DAPI: 4',6-diamidino-2-phenylindole).

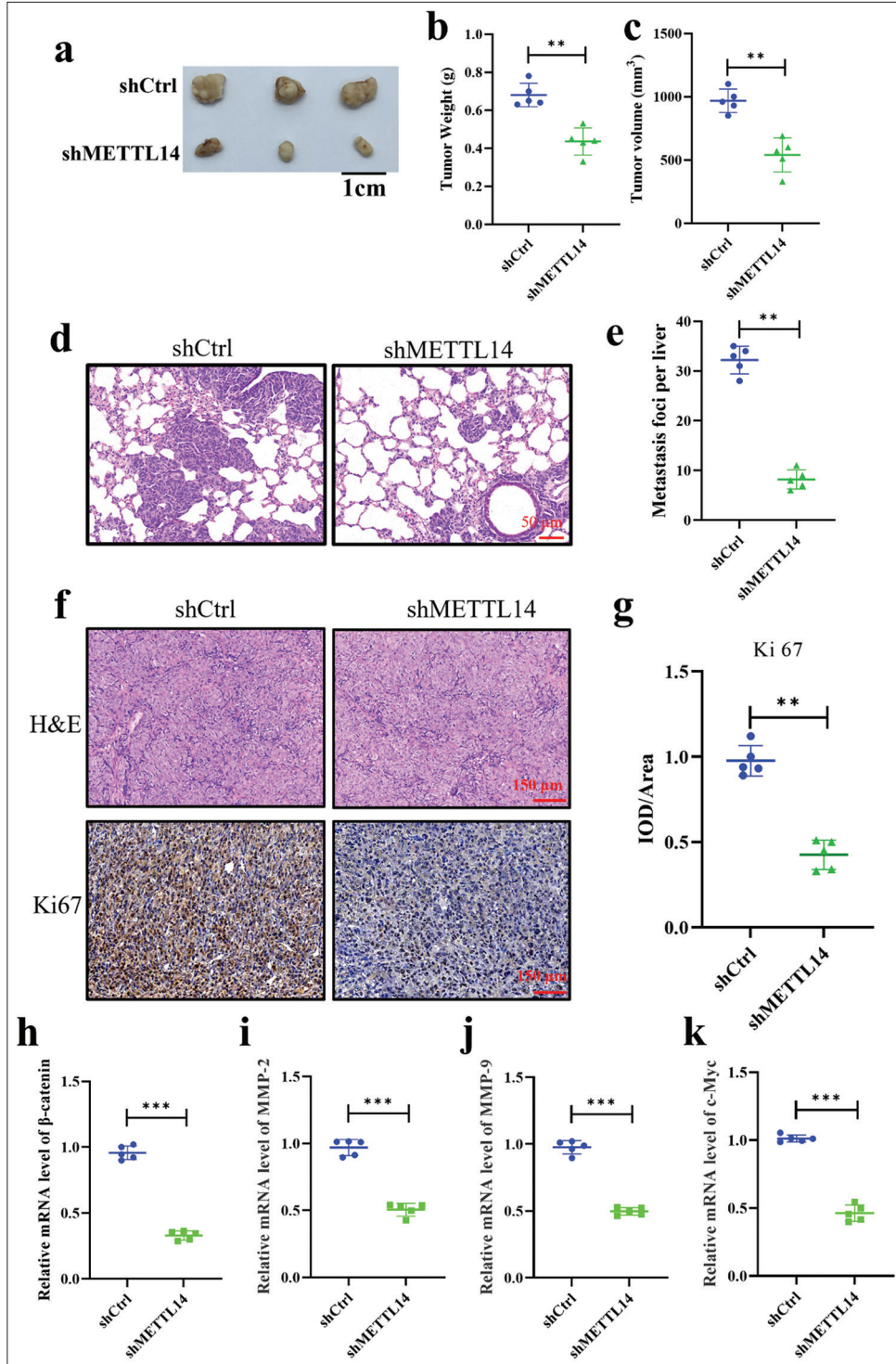
in the OE-NC group at 96 h ( $P < 0.001$ ), and this effect was reversely inhibited by 10058-F4 ( $P < 0.01$ ). Figure 4b demonstrates that the cell proliferation of SW1990 increased at 96 h by OE-METTL14 transfection ( $P < 0.01$ ) and reversely decreased by 10058-F4 ( $P < 0.01$ ). As shown in Figure 4c-e, the increased colony formation ability of OE-METTL14-transfected cells ( $P < 0.001$ ) was reversed by 10058-F4 ( $P < 0.001$ ). In the results of wound scratch healing assay [Figure 4f-h], the increase in cell migration ability in OE-METTL14-transfected cells ( $P < 0.001$ ) was reversed by 10058-F4 ( $P < 0.001$ ). In Figure 4i-k, 10058-F4 treatment also increased the apoptotic cell rate in the OE-METTL14-transfected group ( $P < 0.01$ ). All these results proved that OE-METTL14 upregulated METTL14 expression and promoted the proliferation of pancreatic cancer cell lines PANC-1 and SW1990 by upregulating c-Myc. Otherwise, the

effect of upregulating METTL14 on promoting pancreatic cancer cell proliferation could not be fulfilled.

#### METTL14 promoted tumor growth and metastasis of PANC-1 *in vivo*

To investigate the potential of METTL14 in promoting tumor growth *in vivo*, we generated a stable METTL14 knock down PANC-1 cell line (shMETTL14) and a corresponding negative control (shCtrl) cell line. After 35 days of observation, xenograft tumor volume and weight decreased significantly after silencing METTL14 [Figure 5a-c] ( $P < 0.01$ ). The METTL14 knockdown group had significantly inhibited PANC-1 cell metastases to the lung [Figure 5d and e], ( $P < 0.01$ ). Figure 5f and g revealed a downregulation of Ki-67 in the METTL14 knockdown tumors compared with the





**Figure 5:** METTL14 promoted tumor growth and metastasis of PANC-1 *in vivo*. (a-c) Difference between tumor size and weight after METTL14 knockdown. (d-e) The lung metastasis between METTL14 knockdown and control groups. Objective: 400×. (f-g) H&E and IHC of Ki-67 expression between METTL14 knockdown and control groups. Objective: 400×. (h-k) messenger RNA level of β-catenin, MMP-2, MMP-9, and c-Myc. *n* = 5, \*\**P* < 0.01, \*\*\**P* < 0.001. (H&E: Hematoxylin-eosin, c-Myc: Myc proto-oncogene, MMP: Matrix metalloproteinase).



controls ( $P < 0.01$ ). In addition, shCtrl cells were loosely arranged with gaps. After silencing METTL14, the cells were tightly arranged. Finally, the mRNA expression of  $\beta$ -catenin, MMP-2, MMP-9, and c-Myc was determined by qRT-PCR [Figure 5h-k]. The results of animal experiments were similar to those of PANC-1 cells. The mRNA expression levels of MMP-2, MMP-9,  $\beta$ -catenin, and c-Myc significantly decreased after silencing METTL14 ( $P < 0.001$ ).

## DISCUSSION

The proliferation of pancreatic cancer cells is a critical aspect in tumor growth and malignancy. When pancreatic cancer occurs, the cancer cells will proliferate and their invasion and anti-apoptotic ability will increase.<sup>[21,22]</sup> Our results demonstrate that METTL14 is preferentially upregulated in pancreatic cancer cell lines. The overexpression of METTL14 in these cancer cell lines significantly enhanced their proliferation and colony-forming capabilities, as evidenced by the CCK8 and colony formation assays. Silencing METTL14 led to a marked reduction in these proliferative activities. The TUNEL assay results also indicated that METTL14 overexpression decreased in pancreatic cancer cells. These observations are consistent with a previous report that METTL14 acts as a promoter of tumorigenesis in pancreatic cancer.<sup>[14]</sup> In addition, the downregulation of METTL14 increased apoptosis and autophagy induced by cisplatin in pancreatic cancer cells.<sup>[23]</sup> We further explored the mechanism of action of METTL14 in pancreatic cancer.

The progression of pancreatic cancer is a dynamic process that involves multiple gene expression, modification, and cytokine activation.<sup>[24-27]</sup> c-Myc and beta-catenin are particularly linked to pancreatic cancer. Previous reports found that pancreatic cancer metastasis is promoted by activating c-Myc transcription.<sup>[28]</sup>  $\beta$ -catenin may be the target of miRNAs involved in pancreatic cancer development.<sup>[29]</sup> A critical observation from this study is the effect of METTL14 on c-Myc signaling and  $\beta$ -catenin. The total expression of  $\beta$ -catenin did not change, but the overexpression of METTL14 led to increased  $\beta$ -catenin expression in the cell nucleus. This translocation of  $\beta$ -catenin into the nucleus is crucial because it facilitates the transcription of c-Myc, MMP-9, and MMP-2, which are associated with cell proliferation and metastasis.<sup>[16,17,30]</sup> The upregulation of these key factors underscores the role of METTL14 in promoting a pro-tumorigenic environment through c-Myc signaling.

Furthermore, this study employed the c-Myc inhibitor 10058-F4 to determine the relationship between METTL14 and c-Myc. Many studies have shown that 10058-F4 is a reliable c-Myc inhibitor.<sup>[31-33]</sup> The inhibition of c-Myc effectively reversed the proliferative advantages conferred by METTL14 overexpression, suggesting that c-Myc is a

pivotal target of METTL14 in the context of pancreatic cancer cell proliferation. This finding indicates c-Myc as a central regulator of cell growth and a common oncogene in pancreatic cancer. We further discussed the role of c-Myc in lung adenocarcinoma by combining METTL14 with c-Myc inhibitors. However, this study has some limitations. This study only used animals and cells for experiments, and the types of experimental samples were not rich enough. In the follow-up in-depth investigation, clinical samples of patients may be used to further prove our conclusions.

This study found that METTL14 alleviates the occurrence and metastasis of lung adenocarcinoma by affecting the expression of c-Myc and activating  $\beta$ -catenin into the nucleus.

## SUMMARY

This research highlights the significant role of METTL14 in the proliferation and development of pancreatic cancer. By enhancing c-Myc expression, METTL14 facilitates a cellular environment conducive to tumor growth and survival. These results not only advance our knowledge of the molecular pathways driving the advancement of prostate cancer but they also point to METTL14 as a possible therapeutic target for the creation of novel therapeutic approaches. Further investigations are warranted to explore the therapeutic implications of targeting METTL14 and its downstream signaling pathways in clinical settings.

## AVAILABILITY OF DATA AND MATERIALS

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## ABBREVIATIONS

METTL14 – Methyltransferase-like 14  
 c-Myc – Myc proto-oncogene  
 MMP – Matrix metalloproteinase  
 GAPDH – Glyceraldehyde-3-phosphate dehydrogenase  
 TUNEL – Terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling  
 DAPI – 4',6-diamidino-2-phenylindole  
 H&E – Hematoxylin-eosin  
 qRT-PCR – Quantitative real-time reverse transcription polymerase chain reaction  
 SDS-PAGE – Sodium dodecyl sulfate polyacrylamide gel electrophoresis  
 CCK-8 – Cell counting kit 8  
 IHC – Immunohistochemical

## AUTHORS CONTRIBUTIONS

JRL and PW: Designed the study; All authors conducted the study; FBW and JDD: Collected and analyzed the data. FL, YYL and YYW: Participated in drafting the manuscript, and all authors contributed to critical revision of the manuscript for important intellectual content. All authors gave final approval of the version to be published. All authors participated fully in the work, take public responsibility for appropriate portions of the content, and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or completeness of any part of the work are appropriately investigated and resolved.

## ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study has been approved by the Medical Ethics Committee of Jincheng General Hospital (Approval No.:2024072301 dated 2024.07.23). Because this article did not include any human experiments, informed consent was not required.

## FUNDING

This research was funded by Natural Science Foundation of Shanxi Province. (202203021221293).

## CONFLICT OF INTEREST

The authors declare no conflict of interest.

## EDITORIAL/PEER REVIEW

To ensure the integrity and highest quality of CytoJournal publications, the review process of this manuscript was conducted under a **double-blind model** (authors are blinded for reviewers and vice versa) through an automatic online system.

## REFERENCES

- Li J, Li Y, Chen C, Guo J, Qiao M, Lyu J. Recent estimates and predictions of 5-year survival rate in patients with pancreatic cancer: A model-based period analysis. *Front Med (Lausanne)* 2022;9:1049136.
- Kolbeinson HM, Chandana S, Wright GP, Chung M. Pancreatic cancer: A review of current treatment and novel therapies. *J Invest Surg* 2023;36:2129884.
- Schizas D, Charalampakis N, Kole C, Economopoulou P, Koustas E, Gkotsis E, *et al.* Immunotherapy for pancreatic cancer: A 2020 update. *Cancer Treat Rev* 2020;86:102016.
- Loveday BP, Lipton L, Thomson BN. Pancreatic cancer: An update on diagnosis and management. *Aust J Gen Pract* 2019;48:826-31.
- Torphy RJ, Fujiwara Y, Schulick RD. Pancreatic cancer treatment: Better, but a long way to go. *Surg Today* 2020;50:1117-25.
- Zheng J, Paniccia A, Zureikat AH. Advances in the surgical treatment of pancreatic cancer. *Surg Pathol Clin* 2022;15:479-90.
- Karunakaran M, Barreto SG. Surgery for pancreatic cancer: Current controversies and challenges. *Future Oncol* 2021;17:5135-62.
- Dong L, Chen C, Zhang Y, Guo P, Wang Z, Li J, *et al.* The loss of RNA N(6)-adenosine methyltransferase Mettl14 in tumor-associated macrophages promotes CD8(+) T cell dysfunction and tumor growth. *Cancer Cell* 2021;39:945-57.e10.
- Chen X, Xu M, Xu X, Zeng K, Liu X, Pan B, *et al.* METTL14-mediated N6-methyladenosine modification of SOX4 mRNA inhibits tumor metastasis in colorectal cancer. *Mol Cancer* 2020;19:106.
- Zhou H, Yin K, Zhang Y, Tian J, Wang S. The RNA m6A writer METTL14 in cancers: Roles, structures, and applications. *Biochim Biophys Acta Rev Cancer* 2021;1876:188609.
- Li HB, Huang G, Tu J, Lv DM, Jin QL, Chen JK, *et al.* METTL14-mediated epitranscriptome modification of MN1 mRNA promote tumorigenicity and all-trans-retinoic acid resistance in osteosarcoma. *EBioMedicine* 2022;82:104142.
- Shi B, Liu WW, Yang K, Jiang GM, Wang H. The role, mechanism, and application of RNA methyltransferase METTL14 in gastrointestinal cancer. *Mol Cancer* 2022;21:163.
- Liu X, Du Y, Huang Z, Qin H, Chen J, Zhao Y. Insights into roles of METTL14 in tumors. *Cell Prolif* 2022;55:e13168.
- Wang M, Liu J, Zhao Y, He R, Xu X, Guo X, *et al.* Upregulation of METTL14 mediates the elevation of PERP mRNA N(6) adenosine methylation promoting the growth and metastasis of pancreatic cancer. *Mol Cancer* 2020;19:130.
- Chen S, Yang C, Wang ZW, Hu JF, Pan JJ, Liao CY, *et al.* CLK1/SRSF5 pathway induces aberrant exon skipping of METTL14 and Cyclin L2 and promotes growth and metastasis of pancreatic cancer. *J Hematol Oncol* 2021;14:60.
- Acebron SP, Karaulanov E, Berger BS, Huang YL, Niehrs C. Mitotic wnt signaling promotes protein stabilization and regulates cell size. *Mol Cell* 2014;54:663-74.
- Gentles AJ, Newman AM, Liu CL, Bratman SV, Feng W, Kim D, *et al.* The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat Med* 2015;21:938-45.
- Ala M. Target c-Myc to treat pancreatic cancer. *Cancer Biol Ther* 2022;23:34-50.
- Xi C, Sun J, Xu X, Wu Y, Kou X, Zhao Y, *et al.* Mettl14-driven senescence-associated secretory phenotype facilitates somatic cell reprogramming. *Stem Cell Reports* 2022;17:1799-809.
- Iaiza A, Tito C, Ianniello Z, Ganci F, Laquintana V, Gallo E, *et al.* METTL3-dependent MALAT1 delocalization drives c-Myc induction in thymic epithelial tumors. *Clin Epigenetics* 2021;13:173.
- Narayanan P, Farghadani R, Nyamathulla S, Rajarajeswaran J, Thirugnanasampandan R, Bhuwaneswari G. Natural quinones induce ROS-mediated apoptosis and inhibit cell migration in PANC-1 human pancreatic cancer cell line. *J Biochem Mol Toxicol* 2022;36:e23008.
- Mostafa H, Behrendt I, Meroño T, González-Domínguez R, Fasshauer M, Rudloff S, *et al.* Plasma anthocyanins and their metabolites reduce *in vitro* migration of pancreatic cancer cells, PANC-1, in a FAK- and NF-κB dependent manner:

- Results from the ATTACH-study a randomized, controlled, crossover trial in healthy subjects. *Biomed Pharmacother* 2023;158:114076.
23. Kong F, Liu X, Zhou Y, Hou X, He J, Li Q, *et al.* Downregulation of METTL14 increases apoptosis and autophagy induced by cisplatin in pancreatic cancer cells. *Int J Biochem Cell Biol* 2020;122:105731.
  24. Klein AP. Pancreatic cancer epidemiology: Understanding the role of lifestyle and inherited risk factors. *Nat Rev Gastroenterol Hepatol* 2021;18:493-502.
  25. Ren B, Cui M, Yang G, Wang H, Feng M, You L, *et al.* Tumor microenvironment participates in metastasis of pancreatic cancer. *Mol Cancer* 2018;17:108.
  26. Zhang Z, Xu Y. FZD7 accelerates hepatic metastases in pancreatic cancer by strengthening EMT and stemness associated with TGF- $\beta$ /SMAD3 signaling. *Mol Med* 2022;28:82.
  27. Balsano R, Kruize Z, Lunardi M, Comandatore A, Barone M, Cavazzoni A, *et al.* Transforming growth factor-beta signaling in cancer-induced cachexia: From molecular pathways to the clinics. *Cells* 2022;11:2671.
  28. Xiong Q, Zhang Y, Xu Y, Yang Y, Zhang Z, Zhou Y, *et al.* tiRNA-Val-CAC-2 interacts with FUBP1 to promote pancreatic cancer metastasis by activating c-MYC transcription. *Oncogene* 2024;43:1274-87.
  29. Zhang WT, Zhang JJ, Shao Q, Wang YK, Jia JP, Qian B, *et al.* FGD5-AS1 is an oncogenic lncRNA in pancreatic cancer and regulates the Wnt/ $\beta$ -catenin signaling pathway via miR-577. *Oncol Rep* 2022;47:21.
  30. Hayat R, Manzoor M, Hussain A. Wnt signaling pathway: A comprehensive review. *Cell Biol Int* 2022;46:863-77.
  31. Luo Y, Yang S, Wu X, Takahashi S, Sun L, Cai J, *et al.* Intestinal MYC modulates obesity-related metabolic dysfunction. *Nat Metab* 2021;3:923-39.
  32. Bashash D, Sayyadi M, Safaroghli-Azar A, Sheikh-Zeineddini N, Riyahi N, Momeny M. Small molecule inhibitor of c-Myc 10058-F4 inhibits proliferation and induces apoptosis in acute leukemia cells, irrespective of PTEN status. *Int J Biochem Cell Biol* 2019;108:7-16.
  33. Lv M, Wang Y, Wu W, Yang S, Zhu H, Hu B, *et al.* C-Myc inhibitor 10058-F4 increases the efficacy of dexamethasone on acute lymphoblastic leukaemia cells. *Mol Med Rep* 2018;18:421-8.

**How to cite this article:** Li J, Wang P, Liu F, Li Y, Wu Y, Wang F, *et al.* Methyltransferase-like 14 promotes the tumorigenesis and proliferation of pancreatic cancer cells through myc proto-oncogene signaling pathway. *CytoJournal*. 2024;21:55. doi: 10.25259/Cytojournal\_105\_2024

HTML of this article is available FREE at:  
[https://dx.doi.org/10.25259/Cytojournal\\_105\\_2024](https://dx.doi.org/10.25259/Cytojournal_105_2024)

The FIRST **Open Access** cytopathology journal

Publish in *CytoJournal* and **RETAIN** your copyright for your intellectual property

**Become Cytopathology Foundation (CF) Member at nominal annual membership cost**

For details visit <https://cytojournal.com/cf-member>

**PubMed indexed**

**FREE** world wide **open access**

**Online processing** with rapid turnaround time.

**Real time** dissemination of time-sensitive technology.

Publishes as many **colored high-resolution images**

Read it, cite it, bookmark it, use RSS feed, & many----



**CYTOJOURNAL**

[www.cytojournal.com](http://www.cytojournal.com)

Peer-reviewed academic cytopathology journal







# NextGen CelBloking™ Kits

**Frustrated with your cell blocks?  
We have a better solution!**

**Nano**

## Nano NextGen CelBloking™

Cell block kit to process single scattered cell specimens and tissue fragments of **any** cellularity.



**PATENT PENDING**



**Pack #1**



**Pack #2**

**Micro**

## Micro NextGen CelBloking™

For cellular specimens (more than 1 ml concentrated specimen with Tissuecrit more than 50%)



**PATENT PENDING**



**Pack #2**

