



Editor-in-Chief:

Miaoqing Zhao, PhD., MD (Shandong First Medical University, Jinan, China

Co-editor-in-Chief:

Lester J. Layfield, MD, (University of Missouri, Columbia, MO, USA)

Founding Editor & Editor-in-chief Emeritus:

Vinod B. Shidham, MD, FIAC, FRCPath (WSU School of Medicine, Detroit, USA)

Research Article

Screening for colorectal cancer: Study on the shedding cells of feces

Xin Wang, MD¹, Xiaoxia Qin, MM², Jian Zhang, MD³, Yankai Zhao, MB⁴, Yingchao Gao, MD³

¹Department of Pathology, Hebei Key Laboratory of Colorectal Cancer Precision Diagnosis and Treatment, The First Hospital of Hebei Medical University, Shijiazhuang, China, ²Department of Clinical Laboratory, The First Hospital of Hebei Medical University, Shijiazhuang, China ³Department of Gastrointestinal Surgery, Hebei Key Laboratory of Colorectal Cancer Precision Diagnosis and Treatment, The First Hospital of Hebei Medical University, Shijiazhuang, China, ⁴Department of Gastrointestinal Surgery, The First Hospital of Hebei Medical University, Shijiazhuang, China.



***Corresponding author:**

Yingchao Gao,
Department of Gastrointestinal Surgery, Hebei Key Laboratory of Colorectal Cancer Precision Diagnosis and Treatment, The First Hospital of Hebei Medical University, Shijiazhuang, China.
gaoyingchao2000@163.com

Received: 01 December 2023
Accepted: 01 February 2024
Published: 25 April 2024

DOI
10.25259/Cytojournal_107_2023

Quick Response Code:



ABSTRACT

Objective: The objective of this study was to explore the enrichment efficiency of an improved fecal exfoliated cell enrichment method and its application in colorectal cancer screening.

Material and Methods: Samples were collected from a cohort of 100 colorectal cancer patients being treated at the First Hospital of Hebei Medical University from January 2021 to June 2022. Patient samples were equally divided between control and experimental groups corresponding to the enrichment method being applied to the fecal exfoliated cells. Samples consisted of natural stool and bowel cleansing enema solution samples. The control group received the traditional three-layer integrated screen method, and the experimental group used nano-Fe₃O₄ folic acid magnetic beads to enrich the fecal exfoliated cells. The morphology of the extracted cells was observed by light microscopy through hematoxylin and eosin staining, and the positive rate of fecal occult blood test (FOBT) and the detection rate of colorectal cancer was compared between the two groups.

Results: The FOBT-positive rates of natural feces and intestinal cleansing liquid in the control group were 74.00% and 90.00%, respectively, and the FOBT-positive rates of natural feces and intestinal cleansing liquid in the experimental group were 76.00% and 92.00%, respectively. The positive FOBT rate was high, and the difference was statistically significant ($P = 0.037$ and $P = 0.029$). The sensitivities of natural fecal exfoliation cytology in the control and experimental groups were 82.00% and 92.00%, respectively. The sensitivity of the experimental group was higher than that of the control group, and the difference was not statistically significant ($P = 0.137$). The sensitivities of the exfoliated cytology examination of the intestinal cleansing liquid in the control and experimental groups were 88.00% and 98.00%, respectively. The sensitivity of the experimental group was significantly higher than that of the control group, and the difference was statistically significant ($P = 0.050$). Cell smear results show that the exfoliated cells collected by the three-layer integrated sieve method are unevenly distributed, with overlapping cells and a large number of impurities blurring the background, seriously affecting the observation of cell morphology. The cell structure is blurred, stained unevenly, and arranged in a disorderly manner. The exfoliated cells collected by the nanofolic acid magnetic bead enrichment method are relatively evenly distributed, with no overlapping of cells in patches. The background is clear, and the morphology of each cell can be clearly observed. The cell structure is relatively clear, stained evenly, and distributed evenly.

Conclusion: In the cytological examination of fecal exfoliation of colorectal cancer, the nano-Fe₃O₄ folic acid magnetic bead enrichment method can enrich more target cells compared with the traditional three-layer integrated screen method, improve the detection rate of colorectal cancer, and ensure the exfoliation. The cell smears are of higher quality, providing a better sample for clinical assessment of the exfoliated cells. Nano-Fe₃O₄ folic acid magnetic beads enrichment method can become a simple, efficient, and relatively safe screening method for colorectal cancer, positively affecting early screening developments and diagnosis of colorectal cancer.

Keywords: Feces, Exfoliated cells, Colorectal cancer, Nanotechnology, Enrichment

INTRODUCTION

Colorectal cancer is a common malignant tumor with a high incidence in the anorectal department. Its early symptoms are not obvious; most of them have developed to the middle and late stages when diagnosed.^[1] The incidence of colorectal cancer ranks third among all malignant tumors in China. The incidence of colorectal cancer has increased year by year recently; the age of those affected is getting younger, and the number of male patients is significantly more than that of females.^[2] The 5-year survival rate of colorectal cancer is only 30%. If colorectal cancer can be detected in time at an early stage and undergo surgical resection, patients can have a good prognosis, and the 5-year survival rate can be significantly improved.^[3] Therefore, the key to improving the survival rate of cancer patients lies in early detection, diagnosis, and treatment, and effective early screening is an essential means for timely intervention and treatment.^[4]

Studies have shown that improving early screening can significantly reduce the mortality rate of colorectal cancer.^[5] The fecal occult blood test (FOBT), barium enema, and colonoscopy are all routine screening methods for colorectal cancer. The FOBT is convenient, but its false-positive and false-negative rates are high, and its sensitivity and specificity are poor.^[6] Although barium enemas and colonoscopies have high sensitivity and specificity, they are difficult to promote as first-line screening due to the invasiveness of the procedures and high costs.^[7] Stool testing is simple and specific, with high patient compliance, and can provide early screening and diagnosis of colorectal cancer.^[8]

It has been found in clinical practice that the efficiency in the purification of exfoliated cells is low due to the large amount of impurities mixed in stool, which makes it challenging to popularize stool testing.^[9] Nano-Fe₃O₄ folic acid magnetic beads are widely used in the cytological examination of pleural and ascites exfoliation. According to reports, nano-Fe₃O₄ folic acid magnetic bead technology can enrich cancer cells exfoliated in the sputum of lung cancer patients and tumor cells in the bladder cancer patients' urine while maintaining the isolated target cells' complete morphology and function. Furthermore, it has the advantages of high sensitivity, high specificity, fast detection speed, and easy operation.^[10] Antigen-antibody magnetic bead immune complexes are often used as a research tool to isolate specific cells when detecting circulating tumor cells. Based on previous studies, this study took 100 patients with colorectal cancer as the research object to analyze the enrichment efficiency of the improved fecal exfoliated cell enrichment method and its application value in colorectal cancer screening by providing reference materials for clinical screening of colorectal cancer.

MATERIAL AND METHODS

Research subjects

One hundred colorectal cancer patients (100 cases) admitted to the First Hospital of Hebei Medical University from January 2021 to June 2022 were divided equally (50/group) between the control (three-layer integrated sieve method) and experimental (nanofolate magnetic bead method) groups. The control group comprised 29 males and 21 females; ages ranged from 30 to 72 years, with an average of 53.50 ± 5.65 years; tumor distribution: 21 cases in the rectum, six cases in the descending colon, eight cases in the ascending colon, five cases in the transverse colon, and ten cases in the sigmoid colon; Tumor Dukes staging: 15 cases at stage A, 13 cases at stage B, and 22 cases at stage C; and degree of tumor differentiation: 24 cases were well differentiated, 16 cases were moderately differentiated, and ten cases were poorly differentiated. The experimental group comprised 31 males and 19 females; ages ranged from 28 to 70 years, with an average of 54.25 ± 4.85 years; tumor distribution: 19 cases in the rectum, eight cases in the descending colon, nine cases in the ascending colon, five cases in the transverse colon, and nine cases in the sigmoid colon; Tumor Dukes staging: 16 cases at stage A, 14 cases at stage B, and 20 cases at stage C; and degree of tumor differentiation: 21 cases were well differentiated, 17 cases were moderately differentiated, and 12 cases were poorly differentiated.

There was no statistically significant difference in demographic data such as gender, age, tumor distribution, tumor Dukes stage, and tumor differentiation degree between the two groups.

Inclusion criteria are as follows: (1) Patients diagnosed with colorectal cancer by colonoscopy biopsy. (2) No operation, radiotherapy, or chemotherapy was performed before the feces collection. Exclusion criteria: (1) Patients with colorectal cancer and other malignant tumors. (2) Patients with various types of viral hepatitis. (3) Those with inflammatory bowel disease or other autoimmune diseases comorbidity. (4) Patients with severe cardiovascular disease. (5) Abnormal function of major organs (heart, spleen, and lung).

Instruments and reagents

Immune Fecal Occult Blood Kit (Wanhua Puman Bioengineering Co., Ltd., Cat. No.: PCS0703). Nano-Fe₃O₄ Folic Acid Magnetic Beads Detection Kit (Thermo Fisher Scientific, America, Cat. No.: A48310). Liquid-based Cell Preservation Solution (USA Thermo Fisher Scientific, Cat. No. 12648010). BCD826 bowel cleanser (Shanghai Tiancheng Biotechnology Co., Ltd.). Centrifuge (Beijing Liuyi Instrument Factory). SHZ-88A water bath constant temperature incubator (Taicang Experimental Equipment

Factory, Jiangsu Province). WJ-T6400 automatic cell slicer (Zhejiang Ningbo Medical Equipment Co., Ltd.). ChemiDoc XRS Chemiluminescence imaging analysis system (Bio-rad, USA). Olympus BX-50 optical microscope (Olympus, Japan).

Sample collection

Natural feces

For both the control and experimental groups, 10 g of feces was collected from each subject and visually inspected. Normal appearance permitted suitability for sampling. A total of 5 g samples were collected from five different areas of the stool using a stool key.

Bowel cleansing liquid

Each patient was given an oral laxative on the morning of a colonoscopy. Before colonoscopy, a cleansing enema was performed, and the fourth flushing was retained, filtered, added to the intestinal cell liquid-based preservation solution, and left at room temperature for 2 h.

Cell enrichment

Three-layer integrated sieve method

Natural feces samples were combined with 30 mL of liquid-based preservation solution and mixed thoroughly with a pipette. The total volume, or 100 mL of preserved bowel cleanser sample, was filtered through a 30-mesh (600 μm) screen. The filtrate was transferred to a feces collection bottle with an integrated filter (40 mL) and centrifuged at 1200 rpm for 10 min at room temperature.

The filtrate was further passed through three layers of sieves (screen with 30, 100, and 200 mesh aperture), and the filter residue was discarded. The resulting cell pellet was suspended in 1 mL of liquid-based extraction solution, mixed using a pipette, transferred into the liquid-based thin-layer preparation layer of the automatic cell slicer, and centrifuged at 1000 rpm for 5 min at room temperature. The resulting cell smear was fixed in 95% ethanol for 10 min.

Nanofolate magnetic bead enrichment method

Natural feces samples were combined with 50 mL of cell preservation solution and mixed thoroughly with a pipette. The total volume, or 100 mL of preserved bowel cleanser sample, was filtered through a 30-mesh (600 μm) screen. The filtrate was transferred to a centrifuge tube, and the filter residue was discarded.

The filtrate was centrifuged at 1200 rpm for 10 min at room temperature. The supernatant was discarded, and the pellet

was transferred to a conical centrifuge tube containing 10 mL of cell preservation solution. 40 μL of nanofolate magnetic bead suspension was added, mixed, and incubated in a 37°C water bath for 45 min.

The tube containing the nanofolate magnetic beads-cell suspension was subjected to a magnetic field for 15 min, following which the supernatant was transferred to another conical centrifuge tube.

The nanofolate magnetic bead-cell conjugate was washed down and suspended by mixing.

Both the cell suspension and the conjugation supernatant were centrifuged at 1000 rpm for 10 min. Supernatants were aspirated, leaving 1 mL in each sample. These were resuspended and transferred into the liquid-based thin-layer preparation layer of the automatic cell slicer, centrifuged at 1000 rpm for 5 min at room temperature. The resulting cell smear was fixed in 95% ethanol for 10 min.

Hematoxylin-eosin (HE) staining

The fixed cell smears were dehydrated through an alcohol gradient of alcohol solutions. The alcohol was cleared by incubation in xylene before paraffin embedding. 4 μm slices were cut with a microtome, attached to a glass slide, and dried at 45°C. The samples were again dehydrated, washed with water, and stained with hematoxylin solution for 5 min. After washing, nuclear color depth was observed with an optical microscope. Differentiation was performed in a hydrochloric acid ethanol solution for a few seconds. Samples were counterstained with eosin, washed with water, and dehydrated again. Finally, neutral gum was used to seal the slide. The pathological changes of the cells were observed under an optical microscope, and the images were collected for sorting and analysis.

FOBT

Either 10 mg of natural feces or a bowel cleansing liquid sample was combined with 0.5 mL of distilled water. These were then applied to the colloidal gold test strip, and the results were interpreted within 5 min following the manufacturer's instructions.

Statistical analysis

SPSS 23.0 software was used for statistical analysis of the obtained data. The measurement data were expressed as mean \pm standard deviation. The independent sample t-test was used for comparison between groups, and the count data were expressed as a percentage. The χ^2 test was used for comparison between groups. $P < 0.05$ indicates statistical significance.

RESULTS

Comparison of FOBT-positive rates between two groups of patients

The FOBT-positive rates of natural feces and intestinal cleansing liquid in the control group were 74.00% and 90.00%. The FOBT-positive rates of natural feces and intestinal cleansing liquid in the experimental group were 76.00% and 92.00%. The positive rate of FOBT in the two groups of bowel cleansing solution was higher than that of natural feces FOBT, and the difference was statistically significant ($P < 0.05$); [Table 1].

Comparison of cytology results of natural feces exfoliation between two groups of patients

The sensitivities of natural fecal exfoliation cytology in the control and experimental groups were 82.00% and 92.00%, respectively. The sensitivity of the experimental group was higher than that of the control group, and the difference was not statistically significant ($P = 0.137$); [Table 2].

Comparison of exfoliated cytology results of intestinal cleansing fluid between two groups of patients

The sensitivities of the exfoliated cytology examination of the intestinal cleansing fluid in the control and experimental groups were 88.00% and 98.00%, respectively. The sensitivity of the experimental group was significantly higher than that of the control group, and the difference was statistically significant ($P = 0.050$), as shown in Table 3.

Hematoxylin-eosin (HE) staining to observe the collection of exfoliated cells using the three-layer integrated mesh method

Figure 1a-f shows cancer cells of different shapes and sizes with different degrees of proliferation. Six cell smears were randomly selected to observe the exfoliated cells. Cell smear

results show that the exfoliated cells collected by the three-layer integrated sieve method are unevenly distributed, with overlapping cells and a large number of impurities blurring the background, seriously affecting the observation of cell morphology. The cell structure is blurred, stained unevenly, and arranged in a disorderly manner [Figure 1].

HE staining to observe the collection of exfoliated cells using the nanofolate magnetic bead enrichment method

Figure 2a-f shows cancer cells of different shapes and sizes with different degrees of proliferation. Six cell smears were randomly selected to observe the exfoliated cells. The cell smear results showed that the exfoliated cells collected by the nanofolate magnetic bead enrichment method were relatively evenly distributed, with no overlapping of cells in patches. The background was clean, and the morphology of each cell could be clearly observed. The cell structure is relatively clear, stained evenly, and distributed evenly [Figure 2].

DISCUSSION

Epidemiological surveys have revealed the association between colorectal cancer and both lifestyle and environmental factors; notably, a high-calorie, high-protein, and low-fiber diet accounting for the rapid increase in the incidence of colorectal cancer.^[11] Most colorectal cancers develop from polypoid tumors, taking an average of 10 years to develop from adenoma to cancer.^[12] Studies have found that improving early screening of colorectal cancer can significantly reduce the mortality rate of colorectal cancer.^[13] If yearly stool screening is performed and further microscopic examination is conducted, informed by the results, the mortality rate can be reduced by 33% within 10 years.^[14] There are currently many screening methods for colorectal cancer, among which stool testing has been proven to help detect early-stage cancer and adenoma to improve the prognosis of the disease, in conjunction with removing

Groups	Sample	Number of cases	Positive cases (n)	Negative cases (n)	Positive rate (%)
Control group	Natural feces	50	37	13	74.00
	Bowel cleanser	50	45	5	90.00
χ^2 value					4.336
P-value					0.037
Experimental group	Natural feces	50	38	12	76.00
	Bowel cleanser	50	46	4	92.00
χ^2 value					4.762
P-value					0.029

FOBT: Fecal occult blood test

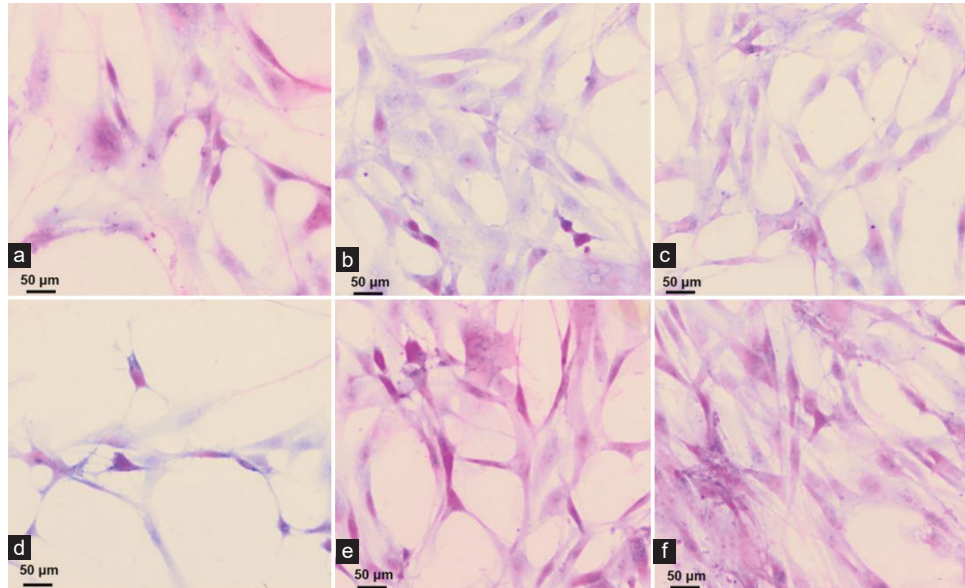


Figure 1: (a-f) Hematoxylin and eosin staining results of fecal exfoliated cells using the three-layer integrated sieve method ($\times 40$, scale bar = 50 μm).

Table 2: Comparison of cytology results of natural feces exfoliation in two groups of patients.

Groups	Number of cases	Positive cases (n)	Negative cases (n)	Sensitivity (%)
Control group	50	41	9	82.00
Experimental group	50	46	4	92.00
χ^2 value				2.210
P-value				0.137

Table 3: Comparison of exfoliated cytology results of intestinal cleansing fluid between two groups of patients.

Groups	Number of cases	Positive cases (n)	Negative cases (n)	Sensitivity (%)
Control group	50	44	6	88.00
Experimental group	50	49	1	98.00
χ^2 value				3.840
P value				0.050

polyps, which can prevent the occurrence of cancer.^[15] The FOBT is easily affected by diet and has low sensitivity to early cancer and adenoma.^[16] There are some limitations with the magnetic bead technology, including the inability to discriminate immunological marker expression by tumors and normal tissues, resulting in false-positive or false-negative results, which will affect the development of clinical applications.^[17] In addition, when preparing magnetic beads,

the antigen coating concentration and coating efficiency can vary, and the prepared magnetic beads have different effects in immunoassay, hampering the unification of detection results.^[18] Therefore, it is essential to find safe and effective screening methods to prevent the occurrence of colorectal cancer that endangers the lives and health of patients. In recent years, nanomagnetic beads have attracted much attention in clinical applications due to their advantages of high separation rate, less impact on active cells, and high specificity.^[19] Folic acid magnetic bead technology provides reference materials for screening, diagnosing, and treating colorectal cancer in the clinic.

In this study, the group was coated on the surface of nano- Fe_3O_4 folic acid magnetic beads to make nanofolate magnetic beads with positive/negative charges. Combined with the principle of tumor cell surface specific screening of cancer cells. It is suggested that the bowel cleansing solution in FOBT can improve the sensitivity to a certain extent. In this study, we investigated the results of exfoliated cytology in natural feces and bowel cleansing fluid of patients with colorectal cancer and found that the positive rate of FOBT in bowel cleansing fluid in both groups was higher than that in natural feces, but the difference was not significant. In the work presented here, the number of cells isolated through the nanofolate magnetic bead method was 0.91×10^6 , confirming the ability of this protocol to enrich colorectal cancer cells, with good specificity shown by the negligible cell density found in the conjugation supernatant. In the exfoliation cytology test, the sensitivity of the experimental group was higher than that of the control group, irrespective of natural feces samples or intestinal cleansing liquid samples,

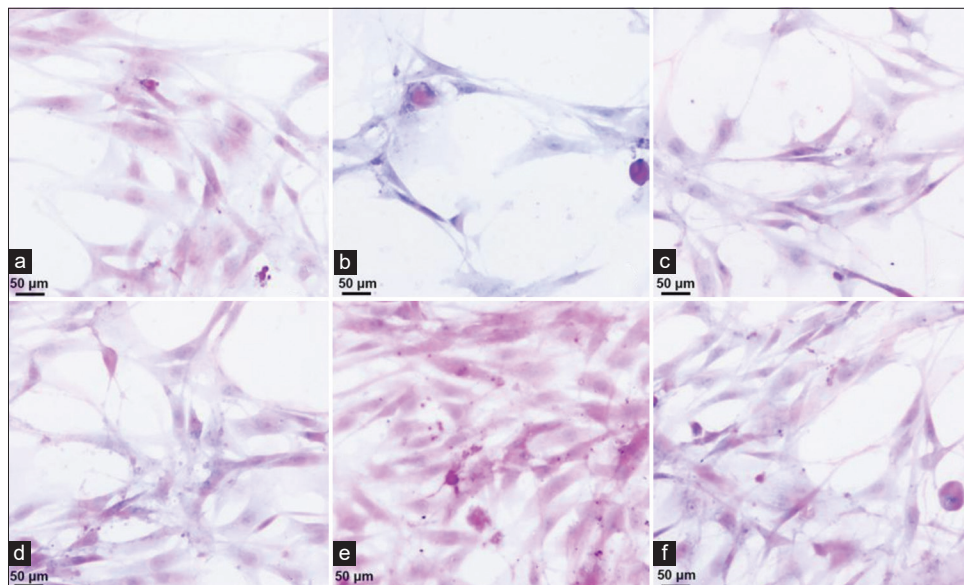


Figure 2: (a-f) Hematoxylin and eosin staining results of fecal exfoliated cells using nanofolate magnetic bead enrichment method ($\times 40$, scale bar = 50 μm).

indicating that the enrichment method of nano folic acid magnetic beads is more effective than the traditional three-layer integrated sieve method.

Enriching fecal exfoliated cells has always been the main factor determining whether the exfoliated cytology method can be promoted in clinical practice.^[20] As early as 1959, the enema method was used to collect exfoliated cells in the intestinal tract to diagnose intestinal tumors. However, its cumbersome operation and low cell enrichment rate make it difficult to popularize.^[21] Studies have used elutriation and silica gel density suspension density gradient centrifugation to extract exfoliated cells from feces. However, due to the anaerobic environment of feces, it is difficult to extract active cells with good morphology.^[22] These limitations have been overcome by a methodology combining whole-feces elution and immunomagnetic beads to extract fecal exfoliated cells, allowing observation of the exfoliated intact cells from feces, and has been applied to the research of other colorectal cancer-related analytical markers.^[23] However, the cell enrichment rate was still low, and a large number of enriched cells were squamous epithelial cells exfoliated from the anal canal.^[24] Nanomagnetic materials are nanoscale particles with a diameter of about 1–100 nm possessing unique magnetic and quantum size effects.^[25] The magnetic nanomaterials can freely penetrate blood vessels and gather under the action of an external magnetic field.^[26] In this study, cell smears were prepared from the enriched cancer cells and atypical cells, allowing specific morphology and structure of exfoliated cells to be observed after fixation and staining. This study found that the nanofolate magnetic bead enrichment method has a higher detection

rate of heterogeneous cells than the traditional three-layer integrated screen method while maintaining the complete morphology and function of the isolated target cells. The quality of the cell smears was higher, with low background, intact cell outline, and even cell distribution, which can aid in better observation of the cell morphology. However, this study has limitations that will be addressed in future work, including a limited sample size, testing a single type of magnetic beads, and using a predetermined concentration of nanofolic acid magnetic beads. At present, there are a variety of early detection methods for colorectal cancer. This study does not claim that one screening method is superior to others; new screening methods are still being explored. It does provide a reference for colorectal cancer screening methods.

SUMMARY

Colorectal cancer is a common anorectal malignancy, and physicians and patients should give early screening sufficient attention. In the cytological examination of fecal exfoliation of colorectal cancer, the nano- Fe_3O_4 folic acid magnetic bead enrichment method can enrich more target cells compared with the traditional three-layer integrated screen method, improve the detection rate of colorectal cancer by maintaining cell quality and morphology which can help doctors better observe the shape of the exfoliated cells. Nano- Fe_3O_4 folic acid magnetic beads enrichment method can become a simple, efficient, and relatively safe screening method for colorectal cancer, which positively affects the development of diagnosis and early screening.

AVAILABILITY OF DATA AND MATERIALS

The data used to support the findings of this study are available from the corresponding author upon request.

ABBREVIATIONS

FOBT - Fecal occult blood test

HE - Hematoxylin-eosin

AUTHOR CONTRIBUTIONS

WX and GY: Experimental design, main experimental implementation, paper writing, XQ: Partial experimental implementation, ZJ: Clinical data collection, YZ: Collected the specimens and carried out some of the experiments, High English Premier League, experimental guidance, paper revision. All authors have reviewed and approved this version. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Ethics committee approval was not required as this was a retrospective cohort study. No intervention was implemented on the patients for the purpose of the study, and no patient-identifiable information was used in the study.

FUNDING

This work was supported by Medical Science Key Research Projects of Hebei China (grant no.20221375 and grant no. 20221412).

CONFLICT OF INTEREST

The authors declare no conflict of interest.

EDITORIAL/PEER REVIEW

To ensure the integrity and highest quality of CytoJournal publications, the review process of this manuscript was conducted under a **double-blind model** (authors are blinded for reviewers and vice versa) through an automatic online system.

REFERENCES

- Patel SG, Karlitz JJ, Yen T, Lieu CH, Boland CR. The rising tide of early-onset colorectal cancer: A comprehensive review of epidemiology, clinical features, biology, risk factors, prevention, and early detection. *Lancet Gastroenterol Hepatol* 2022;7:262-74.
- Baidoun F, Elshiwly KM, Elkeraiya Y, Merjaneh Z, Khoudari G, Sarmini MT, *et al.* Colorectal cancer epidemiology: Recent trends and impact on outcomes. *Curr Drug Targets* 2021;22:998-1009.
- Fan A, Wang B, Wang X, Nie Y, Fan D, Zhao X, *et al.* Immunotherapy in colorectal cancer: Current achievements and future perspective. *Int J Biol Sci* 2021;17:3837-49.
- Wong SH, Yu J. Gut microbiota in colorectal cancer: Mechanisms of action and clinical applications. *Nat Rev Gastroenterol Hepatol* 2019;16:690-704.
- Bretthauer M, Løberg M, Wieszczy P, Kalager M, Emilsson L, Garborg K, *et al.* Effect of colonoscopy screening on risks of colorectal cancer and related death. *N Engl J Med* 2022;387:1547-56.
- Wu Z, Li Y, Zhang Y, Hu H, Wu T, Liu S, *et al.* Colorectal cancer screening methods and molecular markers for early detection. *Technol Cancer Res Treat* 2020;19:1533033820980426.
- Burnett-Hartman AN, Lee JK, Demb J, Gupta S. An update on the epidemiology, molecular characterization, diagnosis, and screening strategies for early-onset colorectal cancer. *Gastroenterology* 2021;160:1041-9.
- Clebak KT, Nickolich S, Mendez-Miller M. Multitarget stool DNA testing (Cologuard) for colorectal cancer screening. *Am Fam Physician* 2022;105:198-200.
- Pei B, Zhang Z, Sun J, Qi X, Cui Q, Yan Y, *et al.* Functionalized ferroferric oxide nanomagnetic beads for extraction of nucleic acid and its application in early screening of colorectal cancer. *J Healthc Eng* 2021;2021:5230666.
- Jin K, Ren C, Liu Y, Lan H, Wang Z. An update on colorectal cancer microenvironment, epigenetic and immunotherapy. *Int Immunopharmacol* 2020;89:107041.
- Picard É, Verschoor CP, Grace WK, Pawelec G. Relationships between immune landscapes, genetic subtypes and responses to immunotherapy in colorectal cancer. *Front Immunol* 2020;11:369.
- Chen H, Li N, Ren J, Feng X, Lyu Z, Wei L, *et al.* Participation and yield of a population-based colorectal cancer screening programme in China. *Gut* 2018;68:1450-7.
- Randel KR, Schult AL, Botteri E, Hoff G, Bretthauer M, Ursin G, *et al.* Colorectal cancer screening with repeated fecal immunochemical test versus sigmoidoscopy: Baseline results from a randomized trial. *Gastroenterology* 2021;160:1085-96.e5.
- Buskermolen M, Cenin DR, Helsingen LM, Guyatt G, Vandvik PO, Haug U, *et al.* Colorectal cancer screening with faecal immunochemical testing, sigmoidoscopy or colonoscopy: A microsimulation modelling study. *BMJ* 2019;367:l5383.
- Klimkiewicz P. USPSTF colorectal cancer screening update 2021. *Nurse Pract* 2022;47:37-42.
- Meklin J, Syrjänen K, Eskelinen M. Colorectal cancer screening with traditional and new-generation fecal immunochemical tests: A critical review of fecal occult blood tests. *Anticancer Res* 2020;40:575-81.
- Jafri HS, Mushtaq S, Baig S. Detection of Kras gene in colorectal cancer patients through liquid biopsy: A cost-effective method. *JCPSP J Coll Phys Surg Pak* 2021;31:1174-8.
- Lu X, Tan S, Wu M, Ju H, Liang X, Li P. Evaluation of a new magnetic bead as an integrated platform for systematic CTC recognition, capture and clinical analysis. *Colloids Surf B Biointerfaces* 2021;199:111542.
- Lin Q, Fang X, Chen H, Weng W, Liu B, Kong J. Dual-modality loop-mediated isothermal amplification for pretreatment-free

- detection of Septin9 methylated DNA in colorectal cancer. *Mikrochim Acta* 2021;188:307.
20. Nakamoto T, Koyama F, Kuge H, Obara S, Ikeda N, Iwasa Y, *et al.* *In vitro* analysis of exfoliated tumor cells in intraluminal lavage samples after colorectal endoscopic submucosal dissection. *Int J Colorectal Dis* 2021;37:161-70.
 21. Wang H, Wang C, Guo L, Zheng Y, Hu W, Dong TT, *et al.* Simultaneous determination of short-chain fatty acids in human feces by HPLC with ultraviolet detection following chemical derivatization and solid-phase extraction segmental elution. *J Sep Sci* 2019;42:2500-9.
 22. Zhou X, Zhang Y, Kang K, Mao Y, Yu Y, Yi Q, *et al.* Controllable environment protein corona-disguised immunomagnetic beads for high-performance circulating tumor cell enrichment. *Anal Chem* 2022;94:4650-7.
 23. Tang J, Dun G, Li J, Xu QL, Zhang T, Lan Y, *et al.* Isolation of *Fusobacterium nucleatum* from human feces using immunomagnetic separation coupled with fastidious anaerobe agar. *J Appl Microbiol* 2022;134:lxac058.
 24. Darroudi M, Gholami M, Rezayi M. An overview and bibliometric analysis on the colorectal cancer therapy by magnetic functionalized nanoparticles for the responsive and targeted drug delivery. *J Nanobiotechnol* 2021;19:399.
 25. Wu K, Su D, Liu J, Saha R, Wang JP. Magnetic nanoparticles in nanomedicine: A review of recent advances. *Nanotechnology* 2019;30:502003.
 26. Li L, Ouyang Y, Hu Y. Research trends on nanomaterials in gastric cancer: A bibliometric analysis from 2004 to 2023. *J Nanobiotechnol* 2023;21:248.

How to cite this article: Wang X, Qin X, Zhang J, Zhao Y, Gao Y. Screening for colorectal cancer: Study on the shedding cells of feces. *CytoJournal*. 2024;21:16. doi: 10.25259/Cytojournal_107_2023

HTML of this article is available FREE at:
https://dx.doi.org/10.25259/Cytojournal_107_2023

The FIRST **Open Access** cytopathology journal

Publish in *CytoJournal* and **RETAIN** your *copyright* for your intellectual property

Become Cytopathology Foundation (CF) Member at nominal annual membership cost

For details visit <https://cytojournal.com/cf-member>

PubMed indexed

FREE world wide **open access**

Online processing with rapid turnaround time.

Real time dissemination of time-sensitive technology.

Publishes as many **colored high-resolution images**

Read it, cite it, bookmark it, use RSS feed, & many----



CYTOJOURNAL

www.cytojournal.com

Peer-reviewed academic cytopathology journal





NextGen CelBloking™ Kits

**Frustrated with your cell blocks?
We have a better solution!**

Nano

Nano NextGen CelBloking™

Cell block kit to process single scattered cell specimens and tissue fragments of **any** cellularity.



PATENT PENDING



Pack #1



Pack #2

Micro

Micro NextGen CelBloking™

For cellular specimens (more than 1 ml concentrated specimen with Tissuecrit more than 50%)



PATENT PENDING



Pack #1



Pack #2