



Research Article

Diagnostic analysis of pleural fluid cell blocks using relevant immunohistochemical markers in clinically suspicious cases of malignancy

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ABSTRACT

Objective: Thoracentesis is a less invasive and expensive procedure than pleural biopsy, enabling sample collection for cytological evaluation. Cell blocks prepared from these samples provide histopathological diagnoses; further, immunohistochemistry (IHC) can be performed for subtyping malignancies, guiding the clinician in choosing the appropriate treatment modality for the patient. We aimed to determine the various histological subtypes of malignancies using cell block IHC and compare the cell block and conventional cytology methods for diagnosing malignant pleural effusion.

Material and Methods: All pleural fluid samples from the clinically suspicious cases of malignancy collected at the Department of Pathology, Government Medical College Kottayam, India, during 18 months, except duplicate and inadequate samples, were included in this prospective observational study. IHC was performed on cell blocks that were positive for malignancy.

Results: This study analyzed 630 samples, of which 121 cell blocks demonstrated the presence of malignancy. Overall, 80%, 13%, and 7% of cases were negative, suspicious, and positive for malignancy based on conventional cytology, and 81%, 0%, and 19% were negative, suspicious, and positive for malignancy based on cell blocks, respectively. Among all malignancies, adenocarcinomas from the lung (56%) were the most common, followed by adenocarcinomas from the breast (6%), squamous cell carcinomas (5%), and adenocarcinomas from the ovary (3%). Cell blocks helped detect more malignancies (19%) than conventional cytology (7%), despite a moderate agreement between both methods.

Conclusion: Adenocarcinomas were the most common cause of malignant pleural effusions, and the most frequent site of origin was the lungs. Cell blocks helped diagnose more malignancies than conventional cytology.

Keywords: Pleural fluid, Conventional cytology, Cell block, Immunohistochemistry, Adenocarcinoma

INTRODUCTION

It is crucial to diagnose cancers at an early stage to provide prompt intervention to patients.^[1,2] Pleural tap or thoracentesis is a less invasive and less expensive procedure than pleural biopsy. Notably, a pleural biopsy has to be performed using a needle under ultrasound or computed tomography guidance under local anesthesia or by thoracoscopy or open surgery under general anesthesia. In contrast, cytological examination of pleural fluids is simple, easy, and inexpensive for detecting malignancies.^[3,4] In addition, an effusion represents cells exfoliated from the entire serosal surface, unlike a focal biopsy from a small area of an extensive serosal surface. Cell blocks prepared from

pleural fluid samples preserve tissue architecture, allowing for histopathological diagnoses of malignancies and additional immunohistochemical or molecular studies to subtype different malignancies.^[5] The subtyping guides the clinician in selecting a suitable treatment option for patients. For example, unlike non-small cell lung cancers, small cell lung cancer requires chemotherapy regardless of the stage of the disease.^[6,7] Moreover, targeted therapy is valuable for treating many oncological diseases (e.g., Epidermal Growth Factor Receptor (EGFR) - or Kirsten Rat Sarcoma Viral Oncogene Homolog (KRAS)-positive lung cancers, Estrogen receptor (ER)/Progesterone receptor (PR)-positive breast cancers, human epidermal growth factor receptor 2 (HER2)-positive breast cancers, etc.).^[8-10] Furthermore, malignant pleural effusions occur because of a primary pleural malignancy (such as malignant mesothelioma) or secondary (metastatic) pleural malignancy (such as adenocarcinoma, small cell carcinoma, myeloma, lymphoma, etc.) and it is challenging to distinguish between reactive mesothelial, adenocarcinoma, and malignant mesothelial cells owing to their overlapping cytological features.^[11,12] Therefore, immunohistochemistry (IHC) is a valuable tool in this regard. Accordingly, this study aimed to determine and characterize various cancers most likely to cause malignant cells in pleural fluids by cell block IHC and determine the mutual diagnostic agreement between conventional cytology and cell block methods.

MATERIAL AND METHODS

This prospective, observational study was conducted at the Department of Pathology (Cytopathology division), Government Medical College Kottayam, India, over 18 months (May 7, 2021–November 6, 2022). The study included pleural fluid samples obtained from clinically suspicious cases of malignancy, whereas duplicate and inadequate samples were excluded. We considered clinically suspicious cases of malignancy as cases with any or combination of the following criteria - the presence of cachexia or significant loss of weight or appetite, rapidly growing swelling or lump or lesion, hard swelling or lump, fixed/non-mobile swelling or lump, non-resolving local site symptoms (such as persistent dyspnea or cough, amenorrhea, bleeding, etc.) after ruling out non-neoplastic causes, elevated serum tumor markers, radiological suggestion or evidence of malignancy, biopsy-proven evidence of cancer at a site other than pleura (such as lung, breast, ovary, endometrium, lymph node, bone marrow, skin, etc.). The clinico-radiological and biopsy details were gathered from patients' medical records. The Scientific Review Committee and Institutional Review Board approved the study. Patient consent was not required for this study, as advised by the Ethics committee, as we had to use only the patients' samples sent to the Department of Pathology by various clinicians who had already got informed consent from patients before performing

thoracentesis. We obtained permission from the department to use the patients' samples for this study.

Pleural fluid samples collected from the clinically suspected cases of malignancy were centrifuged at 2000 rpm for 10 min, the supernatants were discarded, and each sediment was split into three, two of which were transferred onto separate glass slides and one was treated with an acid-alcoholic formalin solution (Formalin: Isopropyl alcohol: Glacial acetic acid= 2: 17: 1). One of the glass slides was stained with Papanicolaou stain, whereas the other stained with Giemsa stain and both were then mounted under coverslips to be later examined under the microscope as conventional cytology slides. Next, centrifugation of the sediment that was mixed with acid-alcoholic formalin solution was performed at 2000 rpm for 10 min, followed by keeping the test tubes aside undisturbed for 4–6 h, after which the supernatants were discarded. Subsequently, the sediments were wrapped in filter papers, placed in separate tissue cassettes, and then placed in a formalin solution overnight. The samples were then processed and embedded in paraffin wax to form tissue-embedded wax blocks (known as "cell blocks"), which were sectioned, stained (using Hematoxylin and Eosin), mounted under coverslips and examined under the microscope.

Conventional cytology or cell block samples were considered positive for malignancy when the microscopic examination revealed atypical cells with a high nuclear-to-cytoplasmic ratio and nuclei containing atypical features such as an irregular nuclear contour, hyperchromasia, intranuclear cytoplasmic inclusions, prominent macronucleoli, or atypical mitosis, either isolated or in clusters (especially if large clusters with >12 cells with molding of the cells or smooth outer borders) or in sheets, which may be present in lacunae (pertaining to cell blocks) and which shows nuclear debris or evidence of necrosis.^[13] The large conglomerate clusters, known as "proliferation spheres" or "cannon balls," may be hollow or solid (without or with stromal cores [papillary]) or a combination (including tubulo-papillary).^[13,14] Hollow proliferation spheres have well-defined internal spaces (glandular-/tubular-/ductal-like or acinar-like) or ill-defined internal spaces (vague glandular-like).^[14,15] Non-cohesive, singly scattered atypical cells or loose clusters would point towards lymphomas or melanomas. If the atypical cells distinctively comprised a second population of cells besides the reactive mesothelial cell population, it was considered metastasis.^[13] The absence of a thin rim of cytoplasm between the nucleus and the cell membrane was considered a diagnostic feature of adenocarcinoma rather than malignant mesothelioma or reactive mesothelial cell proliferation.^[16] Meanwhile, conventional cytology or cell block samples were considered suspicious of malignancy if there were questionable features such as irregular nuclear contour, hyperchromasia, prominent macronucleoli or

atypical mitoses or necrosis. Finally, they were considered negative for malignancy if there were no features such as irregular nuclear contour, hyperchromasia, prominent macronucleoli or atypical mitoses or necrosis.

Moreover, we examined microscopically the histological pattern of arrangement of atypical cells in the cell blocks, which was classified into six groups: cluster (includes simple cluster and solid proliferation sphere), glandular-like hollow proliferation sphere, vague glandular-like hollow proliferation sphere, papillary, sheet, and single-cell patterns. Subsequently, with appropriate controls, we performed IHC on the pleural fluid cell blocks that were positive for malignancy using antibody markers CK7, CK20, TTF1, and calretinin (BioGenex Laboratories, Fremont, CA, USA) and additional IHC markers (BioGenex Laboratories, Fremont, CA, USA) in appropriate cases for subtyping.^[17] The results of IHC marker staining were reported positive if $\geq 5\%$ of the cells (cytoplasm and/or nuclei) were stained, except for ER, which was reported positive if $\geq 1\%$ of the tumor nuclei were stained.^[13,18] We identified various histological subtypes of malignancies based on positive staining by different immunohistochemical markers. In addition, findings from conventional cytology and cell block methods were compared to determine their mutual agreement. Statistical analysis was performed using the SPSS software version 27 (SPSS, Chicago, IL, USA) and kappa statistics.^[19,20] The minimum study sample size (258) to be attained was calculated by using the formula: $n = Zpq/d^2$, where $Z = 1.96$, $p =$ variable with the least proportion, $q = 100 - p$, $d =$ margin of error (taken as 5), and substituting the value of p from the Datta *et al.*^[21] study (metastatic adenocarcinomas from other sites [excluding lung]-21.4%).

RESULTS

Among the pleural fluid samples, most samples were from male patients, and the male-to-female ratio was 1.9:1. Regarding the cell block samples that were positive for malignancy, the male-to-female ratio was 1.3:1. The age of the participants ranged from 45 to 90 years, with a mean age of 55 years. The cases of malignancies diagnosed on cell blocks included patients with a mean age of 60 years.

Overall, 503 (80%) negative, 84 (13%) suspicious, and 43 (7%) positive cases of malignancy were diagnosed on conventional cytology, whereas 509 (81%) negative, 0 (0%) suspicious, and 121 (19%) positive cases of malignancy were diagnosed on cell blocks [Figure 1 and Table 1].

Based on the microscopic examination of cell block samples, the cells of adenocarcinomas exhibited cluster, glandular-like proliferation sphere, and vague glandular-like proliferation sphere patterns. These cells were compared to those of malignant mesothelioma, which demonstrated cluster, vague glandular-like proliferation sphere, papillary, and single-cell patterns. Moreover,

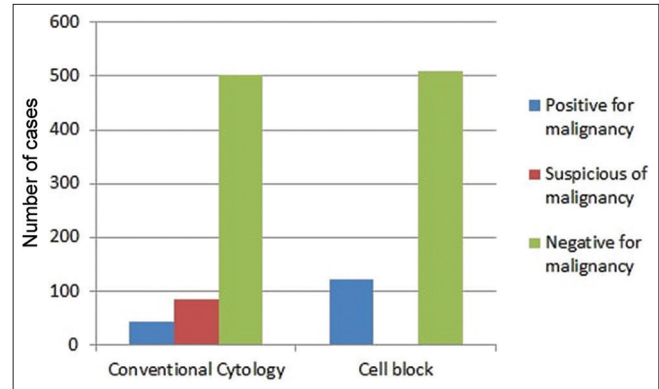


Figure 1: Comparison between conventional cytology and cell blocks.

isolated patterns or various combinations of clusters, sheets, and single cells were observed in other malignancies [Figure 2].

In the present study, immunohistochemical analysis of cell block samples with the initial antibody panel revealed that metastatic adenocarcinomas from the lung [Figure 3] were the most common malignancies (68 cases, 56%), followed by metastatic adenocarcinomas from other sites (21 cases, 17%). In addition, there was 1 case of malignant mesothelioma (1%) and 31 (26%) cases of other malignancies.

Among the metastatic adenocarcinomas from the lung and metastatic adenocarcinomas from other sites, biopsy-proven cases comprised 6 and 5 cases, respectively. None of the other cases were biopsy-proven before the reporting of cell blocks.

Using additional IHC markers, many metastatic adenocarcinomas from other sites were categorized according to their site of origin; moreover, several other malignancies were also diagnosed [Figure 4 and Table 2]. Among metastatic adenocarcinomas from other sites (21 cases, 17%), metastases from the breast were the most common (7 cases, 33%), followed by metastases from the ovary (4 cases, 19%), endometrium (2 cases, 10%), gastrointestinal tract (2 cases, 10%), and urothelium (1 case, 5%). In addition, there were 5 (23%) cases of metastatic adenocarcinomas from unknown sites. Among other malignancies, there were 6 (19%) squamous cell carcinomas, 1 (3.5%) lymphoma, 1 (3.5%) small-cell carcinoma, 1 (3.5%) myeloma, 1 (3.5%) melanoma, 1 (3.5%) thymoma, and 1 (3.5%) embryonal rhabdomyosarcoma.

The diagnostic agreement between conventional cytology and cell block methods was calculated using kappa statistics. Cohen's kappa was found to be 0.54, which indicates a moderate agreement according to the Landis and Koch scale.^[20]

DISCUSSION

Cytological samples are collected by a minimally invasive procedure, unlike biopsy samples. However, in effusion

Conventional cytology				
Cell Block	Negative for malignancy	Suspicious of malignancy	Positive for malignancy	Total
Negative for malignancy	488	21	0	509
Suspicious of malignancy	0	0	0	0
Positive for malignancy	15	63	43	121
Total	503	84	43	630

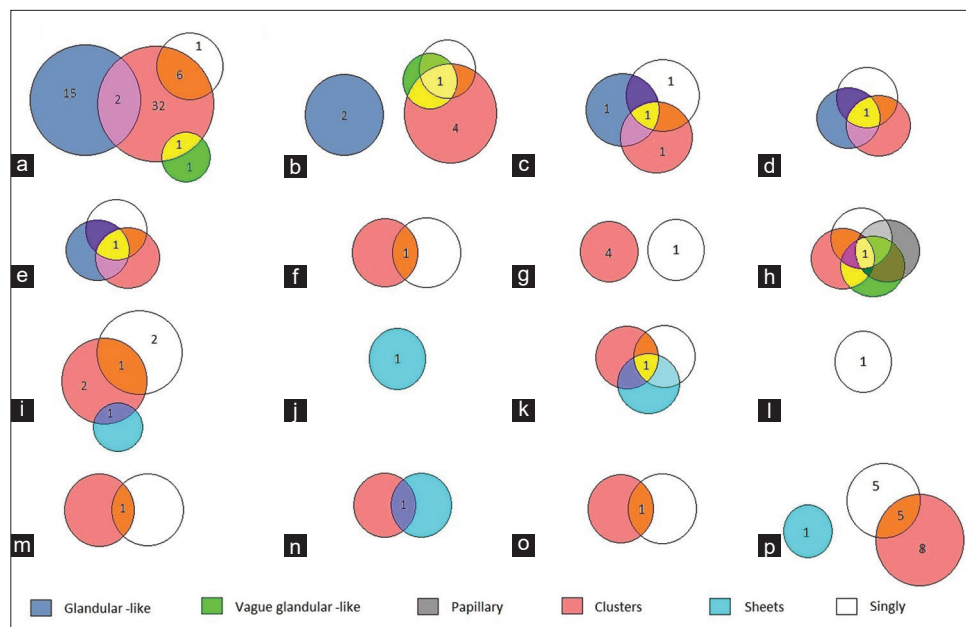


Figure 2: Distribution of patterns of cell arrangement in various malignancies in cell blocks. (a) metastatic adenocarcinoma from the lung, (b) metastatic adenocarcinoma from the breast, (c) metastatic adenocarcinoma from the ovary, (d) metastatic adenocarcinoma from the esophagus, (e) metastatic adenocarcinoma from the colon, (f) metastatic adenocarcinoma from the urothelium, (g) metastatic adenocarcinoma from other sites, (h) malignant mesothelioma, (i) squamous cell carcinoma, (j) lymphoma, (k) myeloma, (l) melanoma, (m) small- cell carcinoma, (n) embryonal rhabdomyosarcoma, (o) thymoma, and (p) unclassified tumors. (Numerals inside the figure indicate the number of cases).

cytology, owing to their broad spectrum of variable morphology, reactive mesothelial cells share many overlapping features with malignant cells of adenocarcinoma or mesothelioma, which poses a significant diagnostic challenge for the cytopathologist. Although there are a few subtle points of differentiation, such as the presence of at least a thin rim of cytoplasm between the cell nucleus and the cell membrane in a mesothelial cell, unlike an adenocarcinoma cell, where the nuclear membrane forms part of the cell outline, not always this is possible for identification.^[16] In this context, immunostaining and/or molecular studies are essential in solving the issue. Cytology smears are not recommended for immunocytochemical staining because of issues related to possible interference with immunoreactivity (due to substitution of formalin or alteration of the processing steps), increased background staining (due to the background

protein-rich material), and lack of opportunity to evaluate coordinate immunoreactivity in the same cells (by Subtractive Coordinate Immunoreactivity Pattern [SCIP] approach). Although immunostaining has been performed successfully on cytology smears (direct smears, cytospin smears, or liquid-based cytology preparations [SurePath™ and ThinPrep™]) following specific protocols, the results are not always readily reproducible by others. Further, collecting cytology samples in weak alcohol fixatives such as Saccomanno Collection fluid, Cytolyt™, PreservCyt™, or CytoRich™ Red would interfere with IHC.^[22-27] This is due to the decreased availability of exposed epitopes that can bind to the antibody. It is here where the utility of cell blocks becomes relevant. This technique has a diagnostic accuracy very close to the gold standard, the pleural biopsy. However, it has been found in some cases that the biopsy specimens have been contaminated with non-

Table 2: Cell block immunohistochemical marker profile of various malignancies.

Metastatic adenocarcinoma from the lung	CK7+, CK20-, TTF1+, Calretinin-, Napsin A+
Metastatic adenocarcinoma from the breast	CK7+, CK20-, TTF1-, Calretinin-, ER+, GATA3+, Vimentin-, Mammaglobin+
Metastatic adenocarcinoma from the ovary	CK7+, CK20-, TTF1-, Calretinin-, ER+, GATA3-, PAX8+, WT1+
Metastatic adenocarcinoma from the endometrium	CK7+, CK20-, TTF1-, Calretinin-, ER+, GATA3+, Vimentin+, CEA-
Metastatic adenocarcinoma from the esophagus ^a	CK7+, CK20+, TTF1-, Calretinin-, CDX2+, SATB2-
Metastatic adenocarcinoma from the colon	CK7-, CK20+, TTF1-, Calretinin-, CDX2+, SATB2+, Beta catenin-nuclear+
Metastatic adenocarcinoma from the urothelium	CK7+, CK20+, TTF1-, Calretinin+, CDX2+, SATB2-, Beta catenin-membranous+, GATA3+
Malignant mesothelioma	CK7+, CK20-, TTF1-, Calretinin+, WT1+, BerEP4-, Claudin4-, BAP1-
Squamous cell carcinoma	CK7-, CK20-, TTF1-, Calretinin-, CK5/6+, p63+, p40+
Lymphoma (T cell type)	CK7-, CK20-, TTF1-, Calretinin-, LCA+, CD3+, CD20 scattered few+
Multiple myeloma	CK7-, CK20-, TTF1-, Calretinin-, CD138+, Kappa+, Lambda-scattered few+
Small cell carcinoma	CK7-, CK20-, TTF1+, Calretinin-, Synaptophysin+, Chromogranin+
Melanoma	CK7-, CK20-, TTF1-, Calretinin-, HMB45+, Melan A+
Thymoma	CK7+, CK20-, TTF1-, Calretinin-, CK5/6+, PAX8+
Embryonal rhabdomyosarcoma	CK7-, CK20-, TTF1-, Calretinin-, LCA-, S100-, CD99-, Desmin+, Myogenin+

^aClinico-radiologic, endoscopic and histopathologic details aided in finding out the site of origin

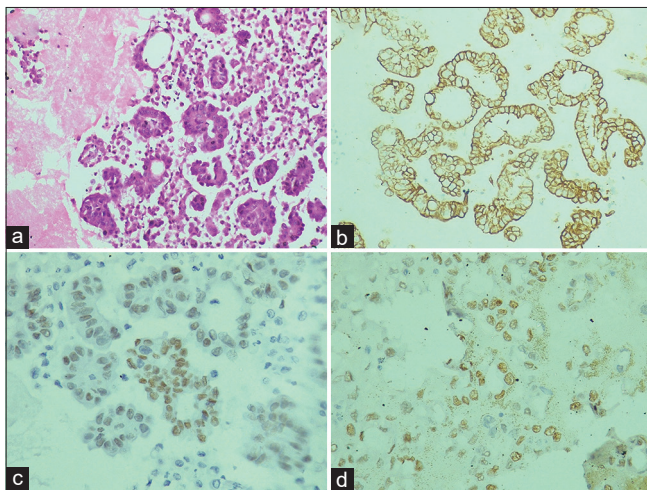


Figure 3: Photomicrographs of the cell block and immunohistochemical sections from metastatic adenocarcinoma from the lung. (a) Cell block section showing hollow proliferation spheres (glandular-like and vague glandular-like), solid proliferation spheres and simple clusters (Hematoxylin and Eosin stain, 10x), which are (b) CK7- cytoplasmic positive (CK7 immunostain, 40x), (c) TTF1- nuclear positive (TTF1 immunostain, 40x), and (d) Napsin A- cytoplasmic positive (Napsin A immunostain, 40x).

neoplastic cells, which can hinder the diagnosis. This type of contamination is less likely in cytology.^[28] Furthermore, unlike biopsy specimens, cell blocks are prepared from cytological fluids obtained by a procedure that causes much less discomfort to the patient. Cell blocks can be prepared by a variety of

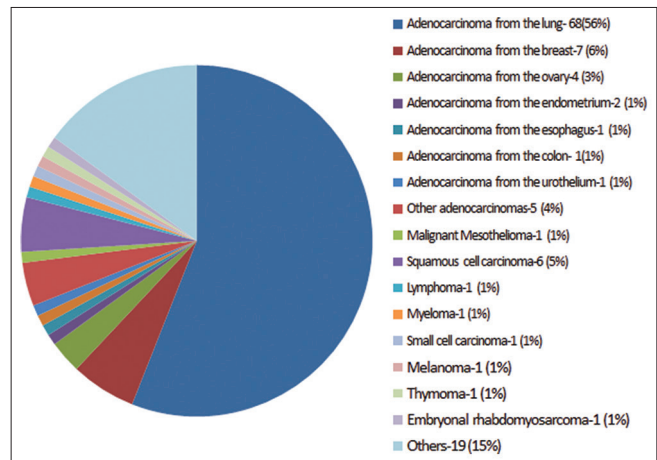


Figure 4: Distribution of diagnoses using cell block immunohistochemistry (IHC).

methods from body fluids. All require prior centrifugation of the fluid sample unless the sample is a clot or from a fat pad, in which case it is processed directly as a cell block. If there is blood contamination, after performing centrifugation, the sediment is treated with BloodLyz™ solution or Ficoll-Hypaque medium before cell block preparation. On the other hand, if there is no blood contamination and the yield of sediment tissue is cellular (>1 mL with Tissucrit [or Cytocrit] >50%), the fixed sediment method may be sufficient for cell block preparation. Techniques such as the plasma thrombin method, agar method, HistoGel™, Collodion bag, Shandon Cytoblock™, Micro NextGen CelBloking™ (Micro-NCGB),

Nano NextGen CelBloking™ (Nano-NCGB), etc. may be used instead for better concentration. Meanwhile, if it is hypocellular (<1 mL with Tissuecrit <50%), Nano-NCGB™ kit should be used for obtaining proper concentration.^[22-24,29] Cell block slides are finally viewed under the microscope to arrive at a histopathological diagnosis. Furthermore, IHC or molecular studies can be performed on cell blocks to reveal the true nature of the lesion as well as identify the subset of patients who will likely benefit from targeted therapy.^[8-10] Cell block IHC performed by the SCIP approach is rewarding as it gives information about the spatial relationship of various cells in the serial sections of the effusion sample, which is absolutely necessary to track the cells when they are stained with an IHC marker as well as decide what next IHC marker has to be performed. In the SCIP approach, all the diagnostic materials are aligned along the potential cutting surface before gelling or embedding the medium for the best yield, which can be achieved by pre-staining with eosin/hematoxylin or by Shidham's protocol using a dark-colored beacon-like AV marker (named after the manufacturer AV BioInnovation LLC, Grosse Ile, MI, USA). The AV marker provides precise monitoring during the section-cutting process, thus facilitating the orientation component of the SCIP approach. NCGB kits already have a built-in AV marker.^[22-24] Recently, dual color SCIP facilitated easy identification of the foreign populations of malignant cells in effusion fluids.^[30] Additionally, special stains may be done on cell-block sections, demonstrating certain organisms, extracellular material or specific histological patterns valuable for diagnosing a particular lesion or neoplasm. Even the frozen section technique has been performed successfully on cell blocks, which allowed rapid diagnosis within a few hours.^[31]

The present study used a large sample size of 630 pleural fluid samples and IHC was performed on 121 cell blocks that showed evidence of malignant cells. In contrast, regarding similar studies^[4,21,32-37] in the relevant literature, most of them used a much lower number of pleural fluid samples and cell block samples for analysis, except for a study by Porcel *et al.*,^[38] which had a sample size (632) comparable to our study, even though they did not perform cell block IHC.

In our study, compared to female patients, the total number of male patients was higher (1.9:1), and this finding agrees with similar studies.^[21,33,36] Further, we demonstrated that as far as cell block samples that were positive for malignancy are concerned, the male-to-female ratio was 1.3:1, which is identical to that observed by Datta *et al.*,^[21] whereas in the studies by Shivakumarswamy *et al.*^[33] and Ranieri *et al.*,^[36] the number of positive cell blocks were higher in female patients. Therefore, the majority of the studies suggest that malignant pleural effusion affects males more than females.

Regarding patients from whom pleural fluid samples were collected in our study, the mean age was 55 years, comparable

to that reported by Datta *et al.*^[21] and Shivakumarswamy *et al.*^[33] Moreover, regarding the patients whose cell block samples were positive for malignancy, their mean age (60) was similar to that reported by Datta *et al.*,^[21] and 56% of them were of age ≥ 60 years. Therefore, studies suggest a higher risk of malignant pleural effusion in individuals of higher age.

Based on conventional cytology, the number of suspicious cases was higher (13%) in the present study than that detected by Shivakumarswamy *et al.*^[33] (8%) and Ranieri *et al.*^[36] (3%). Subsequently, among those 13% suspicious cases, cell block analysis revealed 12% to be positive and 1% to be negative for malignancy. Therefore, the cell block technique eliminated all the suspicious cases that were reported by the conventional cytology method, thereby resolving any issue of diagnostic confusion. Moreover, the cell block method detected more malignancies (19%) than the conventional cytology method (7%), a finding consistent with those of the previous similar studies. The number of malignancies detected on cell blocks by our study (121) lies between those reported by similar studies.^[21,33,36,37]

According to work by Datta *et al.*,^[21] the number of cases of metastatic adenocarcinomas from the lung, metastatic adenocarcinomas from other sites, malignant mesotheliomas, and other malignancies were 15 (53.57%), 6 (21.43%), 0 (0%), and 7 (25%), respectively, and this finding is comparable to that in our study. In contrast, Shivakumarswamy *et al.*^[33] and Ranieri *et al.*^[36] reported that the number of metastatic adenocarcinomas from other sites was more than that from the lung.

In the present study, IHC analysis subtyped 85% of all malignancies. However, owing to the unavailability of some specific IHC markers and/or insufficient residual tissue material, 15% of the cases could not be subtyped and thus remained unclassified malignancies.

This study also found a moderate diagnostic agreement between conventional cytology and cell block methods. However, Shivakumarswamy *et al.*^[33] observed a lower agreement.

Thus, the cell block technique has many distinct advantages of samples being obtained through a less invasive procedure than a pleural biopsy while still providing a histopathological diagnosis, thereby having the provision to do ancillary techniques such as IHC for subtyping malignancies and the capability to detect malignancies that would otherwise be missed on a conventional cytology smear.

SUMMARY

Cell blocks are prepared from pleural fluid samples collected via thoracentesis, a less invasive procedure than a pleural biopsy. In our study, cell block IHC revealed that adenocarcinomas, particularly from the lung, are the major source of malignant cells migrating to the pleural fluid compared to other cancer types. Moreover, cell blocks not

only helped detect more malignancies than conventional cytology but also proved to demonstrate better architectural patterns and be a better platform for performing IHC that altogether aid in arriving at a correct diagnosis.

AVAILABILITY OF DATA AND MATERIALS

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

ABBREVIATIONS

BAP1 - Breast cancer gene 1 associated protein 1
 BerEP4 - Anti-epithelial cell adhesion molecule antibody
 CD3 - Cluster of differentiation 3
 CD20 - Cluster of differentiation 20
 CD99 - Cluster of differentiation 99
 CD138 - Cluster of differentiation 138
 CDX2 - Caudal type homeobox 2
 CEA - Carcinoembryonic antigen
 CK5/6 - Cytokeratin 5/6
 CK 7 - Cytokeratin 7
 CK 20 - Cytokeratin 20
 EGFR - Epidermal growth factor receptor
 ER - Estrogen Receptor
 GATA3 - Guanine adenine thymine adenine binding protein 3
 HMB45 - Human melanoma black 45
 IHC - Immunohistochemistry
 KRAS - Kirsten rat sarcoma viral oncogene homolog
 LCA - Lymphocyte common antigen
 NCGB - NextGen CelBlogging
 PAX8 - Paired box gene 8
 PR - Progesterone Receptor
 SATB2 - Special adenine thymine rich sequence binding protein 2
 SCIP - Subtractive Coordinate Immunoreactivity Pattern
 TTF1 - Thyroid transcription factor 1
 WT1 - Wilms' tumor 1

AUTHOR CONTRIBUTIONS

JKM gathered clinico-radiological details, prepared conventional smears and cell blocks, performed IHC, examined the slides microscopically, and prepared the manuscript. AKK examined the slides microscopically and reviewed the manuscript. GNR reviewed the slides microscopically and reviewed the manuscript. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The Scientific Review Committee and Institutional Review Board approved the study (approval number-30/2021).

Written informed consent was obtained from all the participants prior to the publication of this study.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

EDITORIAL/PEER-REVIEW STATEMENT

To ensure the integrity and highest quality of CytoJournal publications, the review process of this manuscript was conducted under a double-blind model (authors are blinded for reviewers and vice versa) through an automatic online system.

REFERENCES

1. Croswell JM, Ransohoff DF, Kramer BS. Principles of cancer screening: Lessons from history and study design issues. *Semin Oncol* 2010;37:202-15.
2. World Health Organization. Cancer-Screening and Early Detection. Europe: World Health Organization Regional Office for Europe; 2010. Available from: <https://www.who.int/europe/news-room/fact-sheets/item/cancer-screening-and-early-detection-of-cancer> [Last accessed on 2023 Jun 04].
3. Sale SM, Mane VP, Kadam PN. Importance of serosal fluid cytology as an aid to primary diagnosis: A descriptive cross-sectional study. *Natl J Lab Med* 2022;11:PO52-5.
4. Bhanvadia VM, Santwani PM, Vachhani JH. Analysis of diagnostic value of cytological smear method versus cell block method in body fluid cytology: Study of 150 cases. *Ethiop J Health Sci* 2014;24:125-31.
5. Torous VF, Cuda JM, Manucha V, Randolph ML, Shi Q, VandenBussche CJ, *et al.* Cell blocks in cytology: Review of preparation methods, advantages, and limitations. *J Am Soc Cytopathol* 2023;12:77-88.
6. Zappa C, Mousa SA. Non-small cell lung cancer: Current treatment and future advances. *Transl Lung Cancer Res* 2016;5:288-300.
7. Saltos A, Shafique M, Chiappori A. Update on the biology, management, and treatment of small cell lung cancer (SCLC). *Front Oncol* 2020;10:1074.
8. Vormittag-Nocito E, Kumar R, Narayan KD, Chen Z, David O, Behm F, *et al.* Utilization of cytologic cell blocks for targeted sequencing of solid tumors. *Cancer Med* 2023;12:4042-63.
9. Yan L, Rosen N, Arteaga C. Targeted cancer therapies. *Chin J Cancer* 2011;30:1-4.
10. Baudino TA. Targeted cancer therapy: The next generation of cancer treatment. *Curr Drug Discov Technol* 2015;12:3-20.
11. Chowdhuri SR, Fetsch P, Squires J, Kohn E, Filie AC.

- Adenocarcinoma cells in effusion cytology as a diagnostic pitfall with potential impact on clinical management: A case report with brief review of immunomarkers. *Diagn Cytopathol* 2014;42:253-8.
12. Su XY, Li GD, Liu WP, Xie B, Jiang YH. Cytological differential diagnosis among adenocarcinoma, epithelial mesothelioma, and reactive mesothelial cells in serous effusions by immunocytochemistry. *Diagn Cytopathol* 2011;39:900-8.
 13. Cibas ES. Pleural, pericardial, and peritoneal fluids. In: Cibas ES, Ducatman BS, editors. *Diagnostic principles and clinical correlates*. 5th ed. Philadelphia, PA: Elsevier; 2021. p. 141-70.
 14. Khan N, Sherwani RK, Afroz N, Kapoor S. Cytodiagnosis of malignant effusion and determination of primary site. *J Cytol* 2005;22:107-10.
 15. Koss LG, Melamed MR, editors. *Effusions in the presence of cancer*. In: *Koss' diagnostic cytology and its histopathologic bases*. 5th ed. Philadelphia, PA: Lippincott Williams and Wilkins; 2006. p. 949-1022.
 16. Bedrossian CW. Diagnostic problems in serous effusions. *Diagn Cytopathol* 1998;19:131-7.
 17. Tuffaha MS, editor. *Immunohistochemistry in tumor diagnostics*. Cham: Springer; 2018. p. 1-9.
 18. Kim SW, Roh J, Park CS. Immunohistochemistry for pathologists: Protocols, pitfalls, and tips. *J Pathol Transl Med* 2016;50:411-8.
 19. McHugh ML. Interrater reliability: The kappa statistic. *Biochem Med (Zagreb)* 2012;22:276-82.
 20. Landis JR, Koch GG. The measurement of observer agreement for categorical data. *Biometrics* 1977;33:159-74.
 21. Datta P, Saha R, Chakraborty J. A comparative study of conventional cytology and cell block method with immunohistochemistry in the diagnosis of serous effusions. *Trop J Pathol Microbiol* 2020;6:146-54.
 22. Shidham VB, Atkinson BF. *Cytopathologic diagnosis of serous fluids*. 1st ed. China: Elsevier; 2007.
 23. Shidham VB. *CellBlockistry 101: The text book of cell-blocking science*. 1st ed. Grosse Ile, MI: Cytopathology Foundation Inc.; 2021.
 24. Shidham VB, Falzon M. Serous effusions. In: Gray W, Kocjan G, editors. *Diagnostic cytopathology*. 3rd ed. China: Elsevier; 2010. p. 115-78.
 25. Leona AS, Suthipintawong C, Vinyuvat S. Immunostaining of cytologic preparations: A review of technical problems. *Appl Immunohistochem* 1999;7:214-20.
 26. Shidham VB, Lindholm PF, Kajdacsy-Balla A, Chang C, Komorowski R. Methods of cytology smear preparation and fixation: Effect on the immunoreactivity of commonly used anti-cytokeratin antibody AE1/AE3. *Acts Cytol* 2000;44: 1015-22.
 27. Fetsch PA, Abati A. Immunocytochemistry in effusion cytology: A contemporary review. *Cancer Cytopathol* 2001;93: 293-308.
 28. Rani SS, Vamshidhar IS, John NA, John J. Diagnosis of pleural fluid effusions by cell block and pleural biopsy-a comparative study. *J Cytol* 2022;39:169-73.
 29. Processing of single specimen of any cellularity to make a cell block with nano unit. Available from: https://youtube/y29SS1NwO_8 [Last accessed on 2019 Jun 13].
 30. Cihlar KL, Markelova N, Varsegi G, Kumar N, Shidham VB. Two colour immunocytochemistry for evaluation of serous cavity fluids: United States and Canadian Academy of Pathology (USCAP). *Mod Pathol* 2007;20 (supplement 2):A-3804.
 31. Desai F, Korant R. Evaluation of a novel technique of frozen sections of cell blocks and immunocytochemistry protocol for fine needle aspirates and fluid cytology specimens. *Cancer Res Cell Ther* 2022;6:104-9.
 32. Thapar M, Mishra RK, Sharma A, Goyal V, Goyal V. Critical analysis of cell block versus smear examination in effusions. *J Cytol* 2009;26:60-4.
 33. Shivakumarswamy U, Arakeri SU, Karigowdar MH, Yelikar B. Diagnostic utility of the cell block method versus the conventional smear study in pleural fluid cytology. *J Cytol* 2012;29:11-5.
 34. Güldaval F, Anar C, Polat G, Gayaf M, Yavuz MY, Korkmaz A, *et al.* Contribution of cell block obtained by thoracentesis in the diagnosis of malignant pleural effusion. *J Cytol* 2019;36: 205-8.
 35. Shital P, Mirza M, Gondhali G. Pleural fluid "cell block" analysis in malignant pleural effusion: Sensitive, superior over fluid cytology and suitable for immunohistochemistry analysis (IHC), will decrease need for thoracoscopy guided procedures. *Eur Respir J* 2017;50:PA4308.
 36. Ranieri SC, Di Leonardo G, Coletti G, Mas AD, Brancone ML, Crisci R, *et al.* Role of conventional cytology and cell block methods for diagnosis of malignant pleural effusions. *J Xiangya Med* 2020;5:36.
 37. Assawasaksakul T, Boonsarngsuk V, Incharoen P. A comparative study of conventional cytology and cell block method in the diagnosis of pleural effusion. *J Thorac Dis* 2017;9: 3161-7.
 38. Porcel JM, Quirós M, Gatiús S, Bielsa S. Examination of cytological smears and cell blocks of pleural fluid: Complementary diagnostic value for malignant effusions. *Rev Clin Esp (Barc)* 2017;217:144-8.

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